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EP-A- 263 933
EP-A- 0 212 880**BIOCHEM. BIOPHYS. RES. COMMUN., vol. 142, no. 2, 30th January 1987, pages 511-518; R. OIKAWA et al.: "Primary structure of human carcinoembryonic antigen (CEA) deduced from cDNA sequence"****MOL. CELL. BIOL., vol. 7, 1987, page 3221-3230; R. BEAUCHEMIN et al.: "Isolation and characterization of full-length functional cDNA clones for human carcinoembryonic antigen"**(73) Proprietor: **MILES INC.**
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PROC. NATL. ACAD. SCI. USA, vol. 85, September 1988, pages 6959-6963; Y. HINODA et al.: "Molecular cloning of a cDNA coding biliary glycoprotein I: primary structure of a glycoprotein immunologically crossreactive with carcinoembryonic antigen"

GENE, vol. 71, no. 2, November 1988, pages 439-449; B.C. ROONEY et al.: "Molecular cloning of a cDNA for human pregnancy-specific B1-glycoprotein: homology with human carcinoembryonic antigen and related proteins"

Description**BACKGROUND OF THE INVENTION**5 **Field of the Invention**

The present invention concerns nucleic acid sequences which code for carcinoembryonic antigen (CEA) antigen family peptide sequences.

10 **Background Information**

Carcinoembryonic antigen was first described by Gold and Freedman, J. Exp. Med., 121, 439-462, (1965). CEA is characterized as a glycoprotein of approximately 200,000 molecular weight with 50-60% by weight of carbohydrate. CEA is present during normal human fetal development, but only in very low
15 concentration in the normal adult intestinal tract. It is produced and secreted by a number of different tumors.

CEA is a clinically useful tumor marker for the management of colorectal cancer patients. CEA can be measured using sensitive immunoassay methods. When presurgical serum levels of CEA are elevated, a postsurgical drop in serum CEA to the normal range typically indicates successful resection of the tumor.
20 Postsurgical CEA levels that do not return to normal often indicate incomplete resection of the tumor or the presence of additional tumor sites in the patient. After returning to normal levels, subsequent rapid rises in serum CEA levels usually indicate the presence of metastases. Slower postsurgical rises from the normal level are most often interpreted to indicate the presence of new primary tumors not previously detected. Post surgical management of colon cancer patients is thus facilitated by the measurement of CEA.

CEA is a member of an antigen family. Because of this, the immunoassay of CEA by presently available methods is complicated by the fact that CEA is but one of several potentially reactive antigens. There have been at least sixteen CEA-like antigens described in the literature. Since some of these appear to be the same antigen described by different investigators, the actual number of different antigens is somewhat less than this number. Nonetheless, there is a complex array of cross-reactive antigens which
30 can potentially interfere with an immunoassay of the CEA released by tumors. It is known that serum levels of CEA-like antigens are elevated in many non-cancerous conditions such as inflammatory liver diseases and also in smokers. It is important that immunoassays used for the monitoring of cancer patient status not be interfered with by these other CEA-like antigens. Conversely, it is important to be able to distinguish the antigens by immunoassays because of the possibility that different tumor types may preferentially express
35 different forms of CEA. If so, then the ability to reliably measure the different forms of CEA can provide the means to diagnose or more successfully treat different forms of cancer.

The members of the "CEA family" share some antigenic determinants. These common epitopes are not useful in distinguishing the members of the antigen family and antibodies recognizing them are of little use for measuring tumor-specific CEA levels.

40 U.S.P. 3,663,684, entitled "Carcinoembryonic Antigen and Diagnostic Method Using Radioactive Iodine", concerns purification and radioiodination of CEA for use in a RIA.

U.S.P. 3,697,638 describes that CEA is a mixture of antigens (components A and B in this case). U.S.P. 3,697,638 mentions methods for separating and radioiodinating each component and their use in specific RIA's.

45 U.S.P. 3,852,415, entitled "Compositions for Use in Radioimmunoassay, as Substitute for Blood Plasma Extract in Determination of Carcinoembryonic Antigen" relates to the use of a buffer containing EDTA and bovine serum albumin as a substitute for plasma as a diluent for CEA RIA's.

U.S.P. 3,867,363, entitled "Carcinoembryonic Antigens", is directed to the isolation of CEA components A and B, their labelling and use in a RIA.

50 U.S.P. 3,927,193, entitled "Localization of Tumors by Radiolabelled Antibodies", concerns the use of radiolabelled anti-CEA antibodies in whole body tumor imaging.

U.S.P. 3,956,258, entitled "Carcinoembryonic Antigens", relates to the isolation of CEA components A and B.

U.S.P. 4,086,217, entitled "Carcinoembryonic Antigens", is directed to the isolation of CEA components
55 A and B.

U.S.P. 4,140,753, entitled "Diagnostic Method and Reagent", concerns the purification of a CEA isomer called CEA-S1 and its use in a RIA.

U.S.P. 4,145,336, entitled "Carcinoembryonic Antigen Isomer", relates to the antigen CEA-S1.

U.S.P. 4,180,499, entitled "Carcinoembryonic Antigens", describes a process for producing CEA component B.

U.S.P. 4,228,236, entitled "Process of Producing Carcinoembryonic Antigen", is directed to the use of the established cell lines LS-174T and LS-180 or clones or derivatives thereof for the production of CEA.

U.S.P. 4,272,504, entitled "Antibody Adsorbed Support Method for Carcinoembryonic Antigen Assay", concerns two concepts for the radioimmunoassay of CEA. First, U.S.P. 4,272,504 relates to a sample pretreatment in the form of heating to 65 to 85 °C at pH 5 to precipitate and eliminate extraneous protein. Second, it describes the use of a solid phase antibody (either on beads or tubes) as a means to capture analyte and radiolabelled CEA tracer.

U.S.P. 4,299,815, entitled "Carcinoembryonic Antigen Determination", concerns diluting a CEA sample with water and pretreating by heating to a temperature below which precipitation of protein will occur. The pretreated sample is then immunoassayed using RIA, EIA, FIA or chemiluminescent immunoassay.

U.S.P. 4,349,528, entitled "Monoclonal Hybridoma Antibody Specific for High Molecular Weight Carcinoembryonic Antigen", is directed to a monoclonal antibody reacting with 180 kD CEA, but not with other molecular weight forms.

U.S.P. 4,467,031, entitled "Enzyme-Immunoassay for Carcinoembryonic Antigen", relates to a sandwich enzyme immunoassay for CEA in which the first of two anti-CEA monoclonal antibodies is attached to a solid phase and the second monoclonal is conjugated with peroxidase.

U.S.P. 4,489,167, entitled "Methods and Compositions for Cancer Detection", describes that CEA shares an antigenic determinant with alpha-acid glycoprotein (AG), which is a normal component of human serum. The method described therein concerns a solid-phase sandwich enzyme immunoassay using as one antibody an antibody recognizing AG and another antibody recognizing CEA, but not AG.

U.S.P. 4,578,349, entitled "Immunoassay for Carcinoembryonic Antigen (CEA)", is directed to the use of high salt containing buffers as diluents in CEA immunoassays.

EP 113072-A, entitled "Assaying Blood Sample for Carcinoembryonic Antigen - After Removal of Interfering Materials by Incubation with Silica Gel", relates to the removal from a serum of a plasma sample of interfering substances by pretreatment with silica gel. The precleared sample is then subjected to an immunoassay.

EP 102008-A, entitled "Cancer Diagnostics Carcinoembryonic Antigen - Produced from Perchloric Acid Extracts Without Electrophoresis", relates to a procedure for the preparation of CEA from perchloric acid extracts, without the use of an electrophoresis step.

EP 92223-A, entitled "Determination of Carcinoembryonic Antigen in Cytosol or Tissue - for Therapy Control and Early Recognition of Regression", concerns an immunoassay of CEA, not in serum or plasma, but in the cytosol fraction of the tumor tissue itself.

EP 83103759.6, entitled "Cytosole-CEA-Measurement as Predictive Test in Carcinoma, Particularly Mammary Carcinoma", is similar to EP 92223-A.

EP 83303759, entitled "Monoclonal Antibodies Specific to Carcinoembryonic Antigen", relates to the production of "CEA specific" monoclonal antibodies and their use in immunoassays.

WO 84/02983, entitled "Specific CEA-Family Antigens, Antibodies Specific Thereto and Their Methods of Use", is directed to the use of monoclonal antibodies to CEA-meconium (MA)-, and NCA-specific epitopes in immunoassays designed to selectively measure each of these individual components in a sample.

All of the heretofore CEA assays utilize either monoclonal or polyclonal antibodies which are generated by immunizing animals with the intact antigen of choice. None of them address the idea of making sequence specific antibodies for the detection of a unique primary sequence of the various antigens. They do not cover the use of any primary amino acid sequence for the production of antibodies to synthetic peptides or fragments of the natural product. They do not include the concept of using primary amino acid sequences to distinguish the CEA family members. None of them covers the use of DNA or RNA clones for isolating the genes with which to determine the primary sequence.

DEFINITIONSNucleic Acid Abbreviations

5	A	adenine
	G	guanine
	C	cytosine
	T	thymidine
10	U	uracil

Amino Acid Abbreviations:

15	Asp	aspartic acid
	Asn	asparagine
	Thr	threonine
	Ser	serine
20	Glu	glutamic acid
	Gln	glutamine
	Pro	proline
25	Gly	glycine
	Ala	alanine
	Cys	cysteine
30	Val	valine
	Met	methionine
	Ile	isoleucine
35	Leu	leucine
	Tyr	tyrosine
	Phe	phenylalanine
	Trp	tryptophan
40	Lys	lysine
	His	histidine
	Arg	arginine

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Nucleotide - A monomeric unit of DNA or RNA containing a sugar moiety (pentose), a phosphate, and a nitrogenous heterocyclic base. The base is linked to the sugar moiety via the glycosidic carbon (1' carbon of the pentose) and that combination of base and sugar is called a nucleoside. The base characterizes the nucleotide. The four DNA bases are adenine ("A"), guanine ("G"), cytosine ("C"), and thymine ("T"). The four RNA bases are A, G, C and uracil ("U").

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DNA Sequence - A linear array of nucleotides connected one to the other by phosphodiester bonds between the 3' and 5' carbons of adjacent pentoses.

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Functional equivalents - It is well known in the art that in a DNA sequence some nucleotides can be replaced without having an influence on the sequence of the expression product. With respect to the peptide this term means that one or more amino acids which have no function in a particular use can be deleted or replaced by another one.

Codon - A DNA sequence of three nucleotides (a triplet) which encodes through mRNA an amino acid, a translation start signal or a translation termination signal. For example, the nucleotide triplets TTA, TTG,

CTT, CTC, CTA and CTG encode the amino acid leucine ("Leu"), TAG, TAA and TGA are translation stop signals and ATG is a translation start signal.

Reading Frame - The grouping of codons during translation of mRNA into amino acid sequences. During translation, the proper reading frame must be maintained. For example, the sequence
 5 GCTGGTTGTAAG may be translated in three reading frames or phases, each of which affords a different amino acid sequence

GCT GGT TGT AAG - Ala-Gly-Cys-Lys
 10 G CTG GTT GTA AG - Leu-Val-Val
 GC TGG TTG TAA G - Trp-Leu- (STOP) .

Polypeptide - A linear array of amino acids connected one to the other by peptide bonds between the
 15 alpha-amino and carboxy groups of adjacent amino acids.

Genome - The entire DNA of a cell or a virus. It includes inter alia the structural genes coding for the polypeptides of the cell or virus, as well as its operator, promoter and ribosome binding and interaction sequences, including sequences such as the Shine-Dalgarno sequences.

Structural Gene - A DNA sequence which encodes through its template or messenger RNA ("mRNA") a
 20 sequence of amino acids characteristic of a specific polypeptide.

Transcription - The process of producing mRNA from a structural gene.

Translation - The process of producing a polypeptide from mRNA.

Expression - The process undergone by a structural gene to produce a polypeptide. It is a combination of transcription and translation.

25 Plasmid - A non-chromosomal double-stranded DNA sequence comprising an intact "replicon" such that the plasmid is replicated in a host cell. When the plasmid is placed within a unicellular organism, the characteristics of that organism may be changed or transformed as a result of the DNA of the plasmid. For example, a plasmid carrying the gene for tetracycline resistance (Tet^R) transforms a cell previously sensitive to tetracycline into one which is resistant to it. A cell transformed by a plasmid is called a "transformant".

30 Phage or Bacteriophage - Bacterial virus, many of which consist of DNA sequences encapsulated in a protein envelope or coat ("capsid protein").

Cloning Vehicle - A plasmid, phage DNA or other DNA sequence which is capable of replicating in a host cell, which is characterized by one or a small number of endonuclease recognition sites at which such DNA sequences may be cut in a determinable fashion without attendant loss of an essential biological
 35 function of the DNA, e.g., replication, production of coat proteins or loss of promoter or binding sites, and which contains a marker suitable for use in the identification of transformed cells, e.g., tetracycline resistance or ampicillin resistance. A cloning vehicle is often called a vector.

Cloning - The process of obtaining a population of organisms or DNA sequences derived from one such organism or sequence by asexual reproduction.

40 Recombinant DNA Molecule or Hybrid DNA - A molecule consisting of segments of DNA from different genomes which have been joined end-to-end outside of living cells and have the capacity to infect some host cell and be maintained therein.

cDNA Expression Vector - A procaroytic cloning vehicle which also contains sequences of nucleotides that facilitate expression of cDNA sequences in eucaryotic cells. These nucleotides include sequences that
 45 function as eucaryotic promoter, alternative splice sites and polyadenylation signals.

Transformation/Transfection - DNA or RNA is introduced into cells in such a way as to allow gene expression. "Infected" referred to herein concerns the introduction of RNA or DNA by a viral vector into the host.

"Injected" referred to herein concerns the microinjection (use of a small syringe) of DNA into a cell.

50 CEA antigen family (CEA gene family) - a set of genes (gene family) and their products (antigen family) that share nucleotide sequences homologous to partial cDNA LV-7 (CEA-(a)) and as a result of theses similarities also share a subset of their antigenic epitopes. Examples of the CEA antigen family include CEA (= CEA-(b)), transmembrane CEA (TMCEA) = CEA-(c) and normal crossreacting antigen NCA (= CEA-(d)).

SUMMARY OF THE INVENTION

The present invention concerns the following DNA sequences designated as TM-2 (CEA-(e)), TM-3 (CEA-(f)), TM-4 (CEA-(g)), KGCEA1 and KGCEA2, which code for CEA antigen family peptide sequences:

SEQUENCE AND TRANSLATION OF cDNA OF TM-2

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10          10          30          50
CAGCCGTGCTCGAAGCGTTCCTGGAGCCCAAGCTCTCCTCCACAGGTGAAGACAGGGCCA

15          70          90          110
GCAGGAGACACCA˙TGGGGC˙ACCTCTCAGCCCCACTTCACAGAGTGCGTGTACCCTGGCAG
    MetGlyHisLeuSerAlaProLeuHisArgValArgValProTrpGln

20          130          150          170
GGGCTTCTG˙CTCACAGCCTCACTTCTAACC˙TTCTGGAACCCGCCACCACTGCCCAGCTC
GlyLeuLeuLeuThrAlaSerLeuLeuThrPheTrpAsnProProThrThrAlaGlnLeu

25          190          210          230
ACTACTGAATCCATGCCATTCAATGTTGCAGAGGGGAAGGAGGTCTTCTCCTTGTCAC
ThrThrGluSerMetProPheAsnValAlaGluGlyLysGluValLeuLeuLeuValHis

30          250          270          290
AATCTGCCCCAGCAACTTTTTGGCTACAGCTGGTACAAAGGGGAAAGAGTGGATGGCAAC
AsnLeuProGlnGlnLeuPheGlyTyrSerTrpTyrLysGlyGluArgValAspGlyAsn

35          310          330          350
CGTCAAATTGTAGGATATGCAATAGGAACTCAACAAGCTACCCCAAGGCCCGCAAACAGC
ArgGlnIleValGlyTyrAlaIleGlyThrGlnGlnAlaThrProGlyProAlaAsnSer

40          370          390          410
GGTCGAGAGACAATATACCCCAATGCATCCCTGCTGATCCAGAACGTCACCCAGAATGAC
GlyArgGluThrIleTyrProAsnAlaSerLeuLeuIleGlnAsnValThrGlnAsnAsp

45          430          450          470
ACAGGATTCTA˙CACCCTACAAGTCATAAAG˙TCAGATCTTGTGAATGAAGAAGCAACTGGA
ThrGlyPheTyrThrLeuGlnValIleLysSerAspLeuValAsnGluGluAlaThrGly

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490 510 530
CAGTTCCATGTATACCCGGAGCTGCCCAAGCCCTCCATCTCCAGCAACAACCTCCAACCCT
5 GlnPheHisValTyrProGluLeuProLysProSerIleSerSerAsnAsnSerAsnPro

550 570 590
GTGGAGGACAAGGATGCTGTGGCCTTCACCTGTGAACCTGAGACTCAGGACACAACCTAC
10 ValGluAspLysAspAlaValAlaPheThrCysGluProGluThrGlnAspThrThrTyr

610 630 650
CTGTGGTGGATAAAACAATCAGAGCCTCCCGGTGTCAGTCCCAGGCTGCAGCTGTCCAATGGC
15 LeuTrpTrpIleAsnAsnGlnSerLeuProValSerProArgLeuGlnLeuSerAsnGly

670 690 710
AACAGGACCCTCACTCTACTCAGTGTACACAAGGAATGACACAGGACCCTATGAGTGTGAA
20 AsnArgThrLeuThrLeuLeuSerValThrArgAsnAspThrGlyProTyrGluCysGlu

730 750 770
ATACAGAACCAGTGAGTGCGAACCGCAGTGACCCAGTCACCTTGAATGTCACCTATGGC
25 IleGlnAsnProValSerAlaAsnArgSerAspProValThrLeuAsnValThrTyrGly

790 810 830
CCGGACACCCCCACCATTTCCTCCCTTCAGACACCTATTACCGTCCAGGGGCAAACCTCAGC
30 ProAspThrProThrIleSerProSerAspThrTyrTyrArgProGlyAlaAsnLeuSer

850 870 890
CTCTCCTGCTATGCAGCCTCTAACCACCTGCACAGTACTCCTGGCTTATCAATGGAACA
35 LeuSerCysTyrAlaAlaSerAsnProProAlaGlnTyrSerTrpLeuIleAsnGlyThr

910 930 950
TTCCAGCAAAGCACACAAGAGCTCTTTATCCCTAACATCACTGTGAATAATAGTGGATCC
40 PheGlnGlnSerThrGlnGluLeuPheIleProAsnIleThrValAsnAsnSerGlySer

970 990 1010
TATACCTGCCACGCCAATAACTCAGTCACTGGCTGCAACAGGACCACAGTCAAGACGATC
45 TyrThrCysHisAlaAsnAsnSerValThrGlyCysAsnArgThrThrValLysThrIle

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1030 1050 1070
 5 ATAGTCACTGATAATGCTCTACCACAAGAAAATGGCCTCTCACCTGGGGCCATTGCTGGC
 IleValThrAspAsnAlaLeuProGlnGluAsnGlyLeuSerProGlyAlaIleAlaGly

1090 1110 1130
 10 ATTGTGATTGGAGTAGTGGCCCTGGTTGCTCTGATAGCAGTAGCCCTGGCATGTTTCTG
 IleValIleGlyValValAlaLeuValAlaLeuIleAlaValAlaLeuAlaCysPheLeu

1150 1170 1190
 15 CATTTCGGGAAGACCGGCAGGGCAAGCGACCAGCGTGATCTCACAGAGCACAAACCCTCA
 HisPheGlyLysThrGlyArgAlaSerAspGlnArgAspLeuThrGluHisLysProSer

1210 1230 1250
 20 GTCTCCAACCACACTCAGGACCACTCCAATGACCCACCTAACAAGATGAATGAAGTTACT
 ValSerAsnHisThrGlnAspHisSerAsnAspProProAsnLysMetAsnGluValThr

1270 1290 1310
 25 TATTCTACCCTGAACTTTGAAGCCCAGCAACCCACACAACCAACTTCAGCCTCCCCATCC
 TyrSerThrLeuAsnPheGluAlaGlnGlnProThrGlnProThrSerAlaSerProSer

1330 1350 1370
 30 CTAACAGCCACAGAAATAATTTATTTCAGAAAGTAAAAAAGCAGTAATGAAACCTGTCCTGC
 LeuThrAlaThrGluIleIleTyrSerGluValLysLysGln

1390 1410 1430
 35 TCACTGCAGTGCTGATGTATTTCAAGTCTCTCACCCTCATCACTAGGAGATTCTTTCCC

1450 1470 1490
 40 CTGTAGGGTAGAGGGGTGGGGACAGAAACAACCTTCTCCTACTCTTCCTTCCTAATAGGC

1510 1530 1550
 45 ATCTCCAGGCTGCCTGGTCACTGCCCCCTCTCTCAGTGTCATAGATGAAAGTACATTGGG

1570 1590 1610
 50 AGTCTGTAGGAAACCCAACTTCTTGTCATTGAAATTTGGCAAAGCTGACTTTGGGAAAG

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1630 1650 1670
 5 AGGGACCAGAACTTCCCCTCCCTTCCCCTTTTCCCAACCTGGACTTGTTTTAACTTGCC

1690 1710 1730
 10 TGTTTCAGAGCACTCATTCCCTTCCCACCCCCAGTCCTGTCCTATCACTCTAATTTCGGATTT

1750 1770 1790
 15 GCCATAGCCTTGAGGTTATGTCCTTTTCCATTAAAGTACATGTGCCAGGAAACAGCGAGAC

1810 1830 1850
 20 AGAGAAAGTAAACGGCAGTAATGCTTCTCCTATTTCTCCAAAGCCTTGTGTGAAGTAGCA

1870 1890 1910
 25 AAGAGAAGAAAATCAAATATATAACCAATAGTGAAATGCCACAGGTTTGTCCACTGTCAG

1930 1950 1970
 30 GGTTGTCTACCTGTAGGATCAGGGTCTAAGCACCTTGGTGCTTAGCTAGAATACCACCTA

1990 2010 2030
 35 ATCCTTCTGGCAAGCCTGTCTTCAGAGAACCCACTAGAAGCAACTAGGAAAAATCACTTG

2050 2070 2090
 40 CCAAAATCCAAGGCAATTCCCTGATGGAAAATGCAAAAGCACATATATGTTTTAATATCTT

2110 2130 2150
 45 TATGGGCTCTGTTCAAGGCAGTGCTGAGAGGGAGGGTTATAGCTTCAGGAGGGAACCAG

2170 2190 2210
 50 CTTCTGATAAACACAATCTGCTAGGAACTTGGGAAAGGAATCAGAGAGCTGCCCTTCAGC

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2230 2250 2270
 GATTATTTAAATTGTTAAAGAAATACACAATTGGGGTATTGGGATTTTCTCCTTTTCTC
 5 2290 2310 2330
 TGAGACATTCCACCATTTTAATTTTGTAACTGCTTATTTATGTGAAAAGGGTTATTTT
 10 2350 2370 2390
 ACTTAGCTTAGCTATGTCAGCCAATCCGATTGCCTTAGGTGAAAGAAACCACCGAAATCC
 15 2410 2430 2450
 CTCAGGTCCCCTTGGTCAGGAGCCTCTCAAGATTTTTTTTGTGAGAGGCTCCAAATAGAAA
 20 2470 2490 2510
 ATAAGAAAAGGTTTTCTTCATTCATGGCTAGAGCTAGATTTAACTCAGTTTCTAGGCACC
 25 2530 2550 2570
 TCAGACCAATCATCAACTACCATTCTATTCATGTTTGCACCTGTGCATTTTCTGTTTGC
 30 2590 2610 2630
 CCCCATTCACCTTGTGAGGAAACCTTGGCCTCTGCTAAGGTGTATTTGGTCCTTGAGAAG
 35 2650 2670 2690
 TGGGAGCACCTACAGGGACACTATCACTCATGCTGGTGGCATTGTTTACAGCTAGAAAAG
 40 2710 2730 2750
 CTGCACTGGTGCTAATGCCCTTGGGAAATGGGGCTGTGAGGAGGAGGATTATAACTTAG
 45 2770 2790 2810
 GCCTAGCCTCTTTTAAACAGCCTCTGAAAATTTATCTTTTCTTCTATGGGGTCTATAAATGT
 50 2830 2850 2870
 ATCTTTATAAATAAAAAGGAAGGACAGGAGGAAGACAGGCAAAATGTACTTCTCACCAGTCT
 55

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2890 2910 2930
TCTACACAGATGGAATCTCTTTGGGGCTAAGAGAAAGGTTTATTCTATATTGCTTACCT
5
2950 2970 2990
GATCTCATGTTAGGCCTAAGAGGCTTTCTCCAGGAGGATTAGCTTGGAGTTCTCTATACT
10
3010 3030 3050
CAGGTACCTCTTTCAGGGTTTCTAACCCTGACACGGACTGTGCATACTTCCCTCATCC
15
3070 3090 3110
ATGCTGTGCTCTGTTATTTAATTTTTCCTGGCTAAGATCATGTCTGAATTATGTATGAAA
20
3130 3150 3170
ATTATTCTATGTTTTTATAATAAAAAATAATATATCAGACATCGAAAAAAAAA
25
30
35
40
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50
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SEQUENCE AND TRANSLATION OF cDNA OF TM-3

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10 10 30 50
CAGCCGTGCTCGAAGCGTTCCTGGAGCCCAAGCTCTCCTCCACAGGTGAAGACAGGGCCA

15 70 90 110
GCAGGAGACACCATGGGGCACCCTCTCAGCCCCACTTCACAGAGTGCGTGTACCCTGGCAG
MetGlyHisLeuSerAlaProLeuHisArgValArgValProTrpGln

20 130 150 170
GGGCTTCTGCTCACAGCCTCACTTCTAACCTTCTGGAACCCGCCACCCTGCCAGCTC
GlyLeuLeuLeuThrAlaSerLeuLeuThrPheTrpAsnProProThrThrAlaGlnLeu

25 190 210 230
ACTACTGAATCCATGCCATTCAATGTTGCAGAGGGGAAGGAGTTCTTCTCCTTGTCCAC
ThrThrGluSerMetProPheAsnValAlaGluGlyLysGluValLeuLeuLeuValHis

30 250 270 290
AATCTGCCCCAGCAACTTTTTGGCTACAGCTGGTACAAAGGGGAAAGAGTGGATGGCAAC
AsnLeuProGlnGlnLeuPheGlyTyrSerTrpTyrLysGlyGluArgValAspGlyAsn

35 310 330 350
CGTCAAATTGTAGGATATGCAATAGGAACCTCAACAAGCTACCCCAGGGCCCGCAAACAGC
ArgGlnIleValGlyTyrAlaIleGlyThrGlnGlnAlaThrProGlyProAlaAsnSer

40 370 390 410
GGTCGAGAGACAATATACCCCAATGCATCCCTGCTGATCCAGAACGTCACCCAGAATGAC
GlyArgGluThrIleTyrProAsnAlaSerLeuLeuIleGlnAsnValThrGlnAsnAsp

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430 450 470
5 ACAGGATTCTACACCCTACAAGTCATAAAGTCAGATCTTGTGAATGAAGAAGCAACTGGA
ThrGlyPheTyrThrLeuGlnValIleLysSerAspLeuValAsnGluGluAlaThrGly

490 510 530
10 CAGTTCCATGTATACCCGGAGCTGCCCAAGCCCTCCATCTCCAGCAACAACCTCCAACCCT
GlnPheHisValTyrProGluLeuProLysProSerIleSerSerAsnAsnSerAsnPro

550 570 590
15 GTGGAGGACAAGGATGCTGTGGCCTTCACCTGTGAACCTGAGACTCAGGACACAACCTAC
ValGluAspLysAspAlaValAlaPheThrCysGluProGluThrGlnAspThrThrTyr

610 630 650
20 CTGTGGTGGATAAACAATCAGAGCCTCCCGGTCAGTCCCAGGCTGCAGCTGTCCAATGGC
LeuTrpTrpIleAsnAsnGlnSerLeuProValSerProArgLeuGlnLeuSerAsnGly

670 690 710
25 AACAGGACCCTCACTCTACTCAGTGTCAACAAGGAATGACACAGGACCCTATGAGTGTGAA
AsnArgThrLeuThrLeuLeuSerValThrArgAsnAspThrGlyProTyrGluCysGlu

730 750 770
30 ATACAGAACCCAGTGAGTGCGAACCGCAGTGACCCAGTCACCTTGAATGTCACCTATGGC
IleGlnAsnProValSerAlaAsnArgSerAspProValThrLeuAsnValThrTyrGly

790 810 830
35 CCGGACACCCCAACCATTTCCCTTCAGACACCTATTACCGTCCAGGGGCAAACCTCAGC
ProAspThrProThrIleSerProSerAspThrTyrTyrArgProGlyAlaAsnLeuSer

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850 870 890
 CTCTCCTGCTATGCAGCCTCTAACCACCTGCACAGTACTCCTGGCTTATCAATGGAACA
 5 LeuSerCysTyrAlaAlaSerAsnProProAlaGlnTyrSerTrpLeuIleAsnGlyThr
 910 930 950
 TTCCAGCAAAGCACACAAGAGCTCTTTATCCCTAACATCACTGTGAATAATAGTGGATCC
 10 PheGlnGlnSerThrGlnGluLeuPheIleProAsnIleThrValAsnAsnSerGlySer
 970 990 1010
 TATACCTGCCACGCCAATAACTCAGTCACCTGGCTGCAACAGGACCACAGTCAAGACGATC
 15 TyrThrCysHisAlaAsnAsnSerValThrGlyCysAsnArgThrThrValLysThrIle
 1030 1050 1070
 ATAGTCACTGAGCTAAGTCCAGTAGTAGCAAAGCCCCAAATCAAAGCCAGCAAGACCACA
 20 IleValThrGluLeuSerProValValAlaLysProGlnIleLysAlaSerLysThrThr
 1090 1110 1130
 25 GTCACAGGAGATAAGGACTCTGTGAACCTGACCTGCTCCACAAATGACACTGGAATCTCC
 ValThrGlyAspLysAspSerValAsnLeuThrCysSerThrAsnAspThrGlyIleSer
 1150 1170 1190
 30 ATCCGTTGGTTCTTCAAAAACCAGAGTCTCCCGTCCTCGGAGAGGATGAAGCTGTCCAG
 IleArgTrpPhePheLysAsnGlnSerLeuProSerSerGluArgMetLysLeuSerGln
 1210 1230 1250
 35 GGCAACACCACCCTCAGCATAAACCCTGTCAAGAGGGAGGATGCTGGGACGTATTGGTGT
 GlyAsnThrThrLeuSerIleAsnProValLysArgGluAspAlaGlyThrTyrTrpCys

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1270 1290 1310
5 GAGGTCTTCAACCCAATCAGTAAGAACCAAAGCGACCCCATCATGCTGAACGTAAACTAT
GluValPheAsnProIleSerLysAsnGlnSerAspProIleMetLeuAsnValAsnTyr

1330 1350 1370
10 AATGCTCTACCACAAGAAAATGGCCTCTCACCTGGGGCCATTGCTGGCATTGTGATTGGA
AsnAlaLeuProGlnGluAsnGlyLeuSerProGlyAlaIleAlaGlyIleValIleGly

1390 1410 1430
15 GTAGTGGCCCTGGTTGCTCTGATAGCAGTAGCCCTGGCATGTTTTCTGCATTTTCGGGAAG
ValValAlaLeuValAlaLeuIleAlaValAlaLeuAlaCysPheLeuHisPheGlyLys

1450 1470 1490
20 ACCGGCAGCTCAGGACCACTCCAATGACCCACCTAACAAGATGAATGAAGTTACTTATTC
ThrGlySerSerGlyProLeuGln

1510 1530 1550
25 TACCCTGAACTTTGAAGCCCAGCAACCCACACAACCAACTTCAGCCTCCCCATCCCTAAC

1570 1590 1610
30 AGCCACAGAAATAATTTATTCAGAAGTAAAAAGCAGTAATGAAACCTGAAAAAAAAAA

1630
35 AAAAAAAAAA

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SEQUENCE AND TRANSLATION OF cDNA OF TM-4

5 10 30 50
 CAGCCGTGCTCGAAGCGTTCCTGGAGCCCAAGCTCTCCTCCACAGGTGAAGACAGGGCCA

 10 70 90 110
 GCAGGAGACACCATGGGGCACCCTCTCAGCCCCACTTCACAGAGTSCGTGTACCCTGGCAG
 MetGlyHisLeuSerAlaProLeuHisArgValArgValProTrpGln

 15 130 150 170
 GGGCTTCTGCTCACAGCCTCACTTCTAACCTTCTGGAACCCGCCACCCTGCCAGCTC
 GlyLeuLeuLeuThrAlaSerLeuLeuThrPheTrpAsnProProThrThrAlaGlnLeu

 20 190 210 230
 ACTACTGAATCCATGCCATTCAATGTTGCAGAGGGGAAGGAGGTTCTTCTCCTTGTCAC
 ThrThrGluSerMetProPheAsnValAlaGluGlyLysGluValLeuLeuLeuValHis

 25 250 270 290
 AATCTGCCCCAGCAACTTTTGGCTACAGCTGGTACAAAGGGGAAGAGTGGATGGCAAC
 AsnLeuProGlnGlnLeuPheGlyTyrSerTrpTyrLysGlyGluArgValAspGlyAsn

 30 310 330 350
 CGTCAAATTGTAGGATATGCAATAGGAACTCAACAAGCTACCCCAGGGCCCGCAAACAGC
 ArgGlnIleValGlyTyrAlaIleGlyThrGlnGlnAlaThrProGlyProAlaAsnSer

 35 370 390 410
 GGTGAGAGACAATATACCCCAATGCATCCCTGCTGATCCAGAACGTCACCCAGAATGAC
 GlyArgGluThrIleTyrProAsnAlaSerLeuLeuIleGlnAsnValThrGlnAsnAsp

 40 430 450 470
 ACAGGATTCTACACCCTACAAGTCATAAAGTCAGATCTTGTAATGAAGAAGCAACTGGA
 ThrGlyPheTyrThrLeuGlnValIleLysSerAspLeuValAsnGluGluAlaThrGly

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490 510 530
 5 CAGTTCATGTATACCCGGAGCTGCCCAAGCCCTCCATCTCCAGCAACAACCTCCAACCCT
 GlnPheHisValTyrProGluLeuProLysProSerIleSerSerAsnAsnSerAsnPro

550 570 590
 10 GTGGAGGACAAGGATGCTGTGGCCTTCACCTGTGAACCTGAGACTCAGGACACAACCTAC
 ValGluAspLysAspAlaValAlaPheThrCysGluProGluThrGlnAspThrThrTyr

610 630 650
 15 CTGTGGTGGATAAACAATCAGAGCCTCCCGGTCAGTCCCAGGCTGCAGCTGTCCAATGGC
 LeuTrpTrpIleAsnAsnGlnSerLeuProValSerProArgLeuGlnLeuSerAsnGly

670 690 710
 20 AACAGGACCCCTCACTCTACTCAGTGTCAACAAGGAATGACACAGGACCCTATGAGTGTGAA
 AsnArgThrLeuThrLeuLeuSerValThrArgAsnAspThrGlyProTyrGluCysGlu

730 750 770
 25 ATACAGAACCCAGTGAGTGCGAACCGCAGTGACCCAGTCACCTTGAATGTCACCTATGGC
 IleGlnAsnProValSerAlaAsnArgSerAspProValThrLeuAsnValThrTyrGly

790 810 830
 30 CCGGACACCCCCACCATTTCCTTCAGACACCTATTACCGTCCAGGGGCAAACCTCAGC
 ProAspThrProThrIleSerProSerAspThrTyrTyrArgProGlyAlaAsnLeuSer

850 870 890
 35 CTCTCCTGCTATGCAGCCTCTAACCACCTGCACAGTACTCCTGGCTTATCAATGGAACA
 LeuSerCysTyrAlaAlaSerAsnProProAlaGlnTyrSerTrpLeuIleAsnGlyThr

910 930 950
 40 TTCCAGCAAAGCACACAAGAGCTCTTTATCCCTAACATCACTGTGAATAATAGTGGATCC
 PheGlnGlnSerThrGlnGluLeuPheIleProAsnIleThrValAsnAsnSerGlySer

970 990 1010
 45 TATACCTGCCACGCCAATAACTCAGTCACTGGCTGCAACAGGACCACAGTCAAGACGATC
 TyrThrCysHisAlaAsnAsnSerValThrGlyCysAsnArgThrThrValLysThrIle

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1030 1050 1070
 5 ATAGTCACTGATAATGCTCTACCACAAGAAATGGCCTCTCACCTGGGGCCATTGCTGGC
 IleValThrAspAsnAlaLeuProGlnGluAsnGlyLeuSerProGlyAlaIleAlaGly

 1090 1110 1130
 10 ATTGTGATTGGAGTAGTGGCCCTGGTTGCTCTGATAGCAGTAGCCCTGGCATGTTTCTG
 IleValIleGlyValValAlaLeuValAlaLeuIleAlaValAlaLeuAlaCysPheLeu

 1150 1170 1190
 15 CATTTCGGGAAGACCGGCAGCTCAGGACCACTCCAATGACCCACCTAACAAGATGAA TGA
 HisPheGlyLysThrGlySerSerGlyProLeuGln

 1210 1230 1250
 20 AGTTACTTATTCTACCCTGAACTTTGAAGCCCAGCAACCCACACAACCAACTTCAGCCTC

 1270 1290 1310
 25 CCCATCCCTAACAGCCACAGAAATAATTTATTGAGAAGTAAAAAAGCAGTAATGAAACCT

 1330
 30 GAAAAAAAAAAAAAAAAAAAA

The present invention is also directed to a replicable recombinant cloning vehicle ("vector") having an insert comprising a nucleic acid, e.g., DNA, which comprises a base sequence which codes for a CEA peptide or a base sequence hybridizable therewith.

This invention also relates to a cell that is transformed/transfected, infected or injected with the above described replicable recombinant cloning vehicle or nucleic acid hybridizable with the aforementioned cDNA. Thus the invention also concerns the transfection of cells using free nucleic acid, without the use of a cloning vehicle.

Still further, the present invention concerns a polypeptide expressed by the above described transfected, infected or injected cell, which polypeptide exhibits immunological cross-reactivity with a CEA, as well as labelled forms of the polypeptide. The invention also relates to polypeptides having an amino acid sequence, i.e., synthetic peptides, or the expression product of a cell that is transfected, injected, infected with the above described replicable recombinant cloning vehicles, as well as labelled forms thereof. Stated otherwise, the present invention concerns a synthetic peptide having an amino acid sequence corresponding to the entire amino acid sequence or a portion thereof having no less than five amino acids of the aforesaid expression product.

The invention further relates to an antibody preparation specific for the above described polypeptide.

Another aspect of the invention concerns an immunoassay method for detecting CEA or a functional equivalent thereof in a test sample comprising

- (a) contacting the sample with the above described antibody preparation, and
- (b) determining binding thereof to CEA in the sample.

The invention also is directed to a nucleic acid hybridization method for detecting a CEA or a related nucleic acid (DNA or RNA) sample in a test sample comprising

- (a) contacting the test sample with a nucleic acid probe comprising a nucleic acid, which comprises a base sequence which codes for a CEA peptide sequence or a base sequence that is hybridizable therewith, and
- (b) determining the formation of the resultant hybridized probe.

The present invention also concerns a method for detecting the presence of carcinoembryonic antigen or a functional equivalent thereof in an animal or human patient in vivo comprising

a) introducing into said patient a labeled (e.g., a radio-opaque material that can be detected by X-rays, radiolabeled or labeled with paramagnetic materials that can be detected by NMR) antibody preparation according to the present invention and

b) detecting the presence of such antibody preparation in the patient by detecting the label.

In another aspect, the present invention relates to the use of an antibody preparation according to the present invention for therapeutic purposes, namely, attaching to an antibody preparation radionuclides, toxins or other biological effectors to form a complex and introducing an effective amount of such complex into an animal or human patient, e.g., by injection or orally. The antibody complex would attach to CEA in a patient and the radionuclide, toxin or other biological effector would serve to destroy the CEA expressing cell.

BRIEF DESCRIPTION OF THE DRAWINGS

Fig. 1 is a schematic representation of the transmembrane CEA's

DETAILED DESCRIPTION OF THE INVENTION

In the parent application 87111/68, published as EP-A-263 933, applicants described the following CEA's:

	ATCC No.
CEA-(a) partial CEA (pcLV7)	
CEA-(b) full coding CEA (pc 15LV7)	67709
CEA-(c) TM-1 (FL-CEA; pc 19-22)	67710
CEA-(d) NCA (pcBT 20)	67711

In the present application, applicants described the following CEA's:

	ATTC No.
CEA-(e) TM-2 (pc E22)	67712
CEA-(f) TM-3 (pc HT-6)	67708
CEA-(g) TM-4.	

ATCC Nos. 67708, 67709, 67710, 67711 and 67712 were all deposited with the American Type Culture Collection on May 25, 1988.

The sequences for CEA-(a), CEA-(b), CEA-(c) and CEA-(d) are given hereinbelow:

CEA- (a) :

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GG GGT TTA CAC AAC CAC CAC CCC ATC AAA CCC TTC ATC ACC AGC AAC AAC TCC AAC CCC GTG

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GAG GAT GAG GAT GCT GTA GCC TTA ACC TGT GAA CCT GAG ATT CAG AAC ACA ACC TAC CTG

TGG TGG GTA AAT AAT CAG AGC CTC CCG GTC AGT CCC AGG CTG CAG CTG TCC AAT GAC AAC

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AGG ACC CTC ACT CTA CTC AGT GTC ACA AGG AAT GAT GTA GGA CCC TAT GAG TGT GGA ATC

CAG AAC GAA TTA AGT GTT GAC CAC AGC GAC CCA GTC ACC CAG CGA TTC CTC TAT GGC CCA

GAC GAC CCC ACC ATT TCC CCC TCA TAC ACC TAT TAC CGT CCA GGG GTG GAA CCT CAG CCT

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CTC TCC CAT GCA CCC TCT AAC CCA CCT GCA CAG TAT TCT TGG CTG ATT GAT GGG ACC GTC

CAG CAA CAC ACA CAA GAG CTC TTT ATC TCC AAC ATC ACT GAG AAG AAC AGC GCA CTC TAT

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ACC TGC CAG GCC AAT AAC TCA GCC AGT GGC ACA GCA GGA CTA CAG TCA AGA CAA TCA CAG

TCT CTG CCG ATG CCC AAG CCC TCC ATC TCC AGC AAC AAC TCC AAA CCC GTG GAG GAC AAG

GAT CCC TGT GGC CTT CAC TGT GAA CCT GAG GCT CAG AAC ACA ACC TAC CTG TGG TGG GTA

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AAT GGT CAG AGC CTC CCA GTC AGT CCC AGG CTG CAG CTG TCC AAT GGC AAC AGG ACC CTC

ACT CTA TTC AAT GTC ACA AGA AAT GAC GCA AGA GCC TAT GTA TGT GGA ATC CAG AAC TCA

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GTG AGT GCA AAC CCC AGT GAC CCA GTC ACC CTG GAT GTC CTC TAT GGG CCG GAC ACC CCC

ATC ATT TCC CCC CCC CC

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(b)

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C ACC ATG GAG TCT CCC TCG GCC CCT CTC CAC AGA TGG TGC ATC CCC TGG CAG AGG CTC
Met Glu Ser Pro Ser Ala Pro Leu His Arg Trp Cys Ile Pro Trp Gln Arg Leu

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520 530 540 550 560 570
 ACT CAG GAC GCA ACC TAC CTG TGG TGG GTA AAC AAT CAG AGC CTC CCG GTC AGT CCC
 Thr Gln Asp Ala Thr Tyr Leu Trp Trp Val Asn Asn Gln Ser Leu Pro Val Ser Pro
 137 138 139 140 141 142 143 144 145 146 147 148 149 150 151 152 153 154 155

580 590 600 610 620
 AGG CTG CAG CTG TCC AAT GGC AAC AGG ACC CTC ACT CTA TTC AAT GTC ACA AGA AAT
 Arg Leu Gln Leu Ser Asn Gly Asn Arg Thr Leu Thr Leu Phe Asn Val Thr Arg Asn
 156 157 158 159 160 161 162 163 164 165 166 167 168 169 170 171 172 173 174

630 640 650 660 670 680
 GAA CAA GCA AGC TAC AAA TGT GAA ACC CAG AAC CCA GTG AGT GCC AGG CGC AGT GAT
 Glu Gln Ala Ser Tyr Lys Cys Glu Thr Gln Asn Pro Val Ser Ala Arg Arg Ser Asp
 175 176 177 178 179 180 181 182 183 184 185 186 187 188 189 190 191 192 193

690 700 710 720 730 740
 TCA GTC ATC CTG AAT GTC CTC TAT GGC CCG GAT GCC CCC ACC ATT TCC CCT CTA AAC
 Ser Val Ile Leu Asn Val Leu Tyr Gly Pro Asp Ala Pro Thr Ile Ser Pro Leu Asn
 194 195 196 197 198 199 200 201 202 203 204 205 206 207 208 209 210 211 212

750 760 770 780 790
 ACA TCT TAC AGA TCA GGG GAA AAT CTG AAC CTC TCC TGC CAC GCA GCC TCT AAC CCA
 Thr Ser Tyr Arg Ser Gly Glu Asn Leu Asn Leu Ser Cys His Ala Ala Ser Asn Pro
 213 214 215 216 217 218 219 220 221 222 223 224 225 226 227 228 229 230 231

800 810 820 830 840 850
 CCT GCA CAG TAC TCT TGG TTT GTC AAT GGG ACT TTC CAG CAA TCC ACC CAA GAG CTC
 Pro Ala Gln Tyr Ser Trp Phe Val Asn Gly Thr Phe Gln Gln Ser Thr Gln Glu Leu
 232 233 234 235 236 237 238 239 240 241 242 243 244 245 246 247 248 249 250

860 870 880 890 900 910
 TTT ATC CCC AAC ATC ACT GTG AAT AAT AGT GGA TCC TAT ACG TGC CAA GCC CAT AAC
 Phe Ile Pro Asn Ile Thr Val Asn Asn Ser Gly Ser Tyr Thr Cys Gln Ala His Asn
 251 252 253 254 255 256 257 258 259 260 261 262 263 264 265 266 267 268 269

920 930 940 950 960 970
 TCA GAC ACT GGC CTC AAT AGG ACC ACA GTC ACG ACG ATC ACA GTC TAT GCA GAG CCA
 Ser Asp Thr Gly Leu Asn Arg Thr Thr Val Thr Thr Ile Thr Val Tyr Ala Glu Pro
 270 271 272 273 274 275 276 277 278 279 280 281 282 283 284 285 286 287 288

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 980 990 1000 1010 1020
 CCC AAA CCC TTC ATC ACC AGC AAC AAC TCC AAC CCC GTG GAG GAT GAG GAT GCT GTA
 Pro Lys Pro Phe Ile Thr Ser Asn Asn Ser Asn Pro Val Glu Asp Glu Asp Ala Val
 289 290 291 292 293 294 295 296 297 298 299 300 301 302 303 304 305 306 307

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 1030 1040 1050 1060 1070 1080
 GCC TTA ACC TGT GAA CCT GAG ATT CAG AAC ACA ACC TAC CTG TGG TGG GTA AAT AAT
 Ala Leu Thr Cys Glu Pro Glu Ile Gln Asn Thr Thr Tyr Leu Trp Trp Val Asn Asn
 308 309 310 311 312 313 314 315 316 317 318 319 320 321 322 323 324 325 326

15
 1090 1100 1110 1120 1130 1140
 CAG AGC CTC CCG GTC AGT CCC AGG CTG CAG CTG TCC AAT GAC AAC AGG ACC CTC ACT
 Gln Ser Leu Pro Val Ser Pro Arg Leu Gln Leu Ser Asn Asp Asn Arg Thr Leu Thr
 327 328 329 330 331 332 333 334 335 336 337 338 339 340 341 342 343 344 345

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 1150 1160 1170 1180 1190
 CTA CTC AGT GTC ACA AGG AAT GAT GTA GGA CCC TAT GAG TGT GGA ATC CAG AAC GAA
 Leu Leu Ser Val Thr Arg Asn Asp Val Gly Pro Tyr Glu Cys Gly Ile Gln Asn Glu
 346 347 348 349 350 351 352 353 354 355 356 357 358 359 360 361 362 363 364

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 1200 1210 1220 1230 1240 1250
 TTA AGT GTT GAC CAC AGC GAC CCA GTC ATC CTG AAT GTC CTC TAT GGC CCA GAC GAC
 Leu Ser Val Asp His Ser Asp Pro Val Ile Leu Asn Val Leu Tyr Gly Pro Asp Asp
 365 366 367 368 369 370 371 372 373 374 375 376 377 378 379 380 381 382 383

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 1260 1270 1280 1290 1300 1310
 CCC ACC ATT TCC CCC TCA TAC ACC TAT TAC CGT CCA GGG GTG AAC CTC AGC CTC TCC
 Pro Thr Ile Ser Pro Ser Tyr Thr Tyr Tyr Arg Pro Gly Val Asn Leu Ser Leu Ser
 384 385 386 387 388 389 390 391 392 393 394 395 396 397 398 399 400 401 402

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 1320 1330 1340 1350 1360
 TGC CAT GCA GCC TCT AAC CCA CCT GCA CAG TAT TCT TGG CTG ATT GAT GGG AAC ATC
 Cys His Ala Ala Ser Asn Pro Pro Ala Gln Tyr Ser Trp Leu Ile Asp Gly Asn Ile
 403 404 405 406 407 408 409 410 411 412 413 414 415 416 417 418 419 420 421

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 1370 1380 1390 1400 1410 1420
 CAG CAA CAC ACA CAA GAG CTC TTT ATC TCC AAC ATC ACT GAG AAG AAC AGC GGA CTC
 Gln Gln His Thr Gln Glu Leu Phe Ile Ser Asn Ile Thr Glu Lys Asn Ser Gly Leu
 422 423 424 425 426 427 428 429 430 431 432 433 434 435 436 437 438 439 440

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 441 442 443 444 445 446 447 448 449 450 451 452 453 454 455 456 457 458 459 460

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 461 462 463 464 465 466 467 468 469 470 471 472 473 474 475 476 477 478 479 480

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 481 482 483 484 485 486 487 488 489 490 491 492 493 494 495 496 497 498 499 500

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5 1430 1440 1450 1460 1470 1480
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TAT ACC TGC CAG GCC AAT AAC TCA GCC AGT GGC CAC AGC AGG ACT ACA GTC AAG ACA
Tyr Thr Cys Gln Ala Asn Asn Ser Ala Ser Gly His Ser Arg Thr Thr Val Lys Thr
441 442 443 444 445 446 447 448 449 450 451 452 453 454 455 456 457 458 459

10 1490 1500 1510 1520 1530 1540
 • • • • • •
ATC ACA GTC TCT GCG GAC GTG CCC AAG CCC TCC ATC TCC AGC AAC AAC TCC AAA CCC
Ile Thr Val Ser Ala Asp Val Pro Lys Pro Ser Ile Ser Ser Asn Asn Ser Lys Pro
460 461 462 463 464 465 466 467 468 469 470 471 472 473 474 475 476 477 478

15 1550 1560 1570 1580 1590
 • • • • •
GTG GAG GAC AAG GAT GCT GTG GCC TTC ACC TGT GAA CCT GAG GCT CAG AAC ACA ACC
Val Glu Asp Lys Asp Ala Val Ala Phe Thr Cys Glu Pro Glu Ala Gln Asn Thr Thr
479 480 481 482 483 484 485 486 487 488 489 490 491 492 493 494 495 496 497

20 1600 1610 1620 1630 1640 1650
 • • • • • •
TAC CTG TGG TGG GTA AAT GGT CAG AGC CTC CCA GTC AGT CCC AGG CTG CAG CTG TCC
Tyr Leu Trp Trp Val Asn Gly Gln Ser Leu Pro Val Ser Pro Arg Leu Gln Leu Ser
498 499 500 501 502 503 504 505 506 507 508 509 510 511 512 513 514 515 516

25 1660 1670 1680 1690 1700 1710
 • • • • • •
AAT GGC AAC AGG ACC CTC ACT CTA TTC AAT GTC ACA AGA AAT GAC GCA AGA GCC TAT
Asn Gly Asn Arg Thr Leu Thr Leu Phe Asn Val Thr Arg Asn Asp Ala Arg Ala Tyr
517 518 519 520 521 522 523 524 525 526 527 528 529 530 531 532 533 534 535

30 1720 1730 1740 1750 1760
 • • • • •
GTA TGT GGA ATC CAG AAC TCA GTG AGT GCA AAC CGC AGT GAC CCA GTC ACC CTG GAT
Val Cys Gly Ile Gln Asn Ser Val Ser Ala Asn Arg Ser Asp Pro Val Thr Leu Asp
536 537 538 539 540 541 542 543 544 545 546 547 548 549 550 551 552 553 554

35 1770 1780 1790 1800 1810 1820
 • • • • • •
GTC CTC TAT GGG CCG GAC ACC CCC ATC ATT TCC CCC CCA GAC TCG TCT TAC CTT TCG
Val Leu Tyr Gly Pro Asp Thr Pro Ile Ile Ser Pro Pro Asp Ser Ser Tyr Leu Ser
555 556 557 558 559 560 561 562 563 564 565 566 567 568 569 570 571 572 573

40 1830 1840 1850 1860 1870 1880
 • • • • • •
GGA GCG AAC CTC AAC CTC TCC TGC CAC TCG GCC TCT AAC CCA TCC CCG CAG TAT TCT
Gly Ala Asn Leu Asn Leu Ser Cys His Ser Ala Ser Asn Pro Ser Pro Gln Tyr Ser
574 575 576 577 578 579 580 581 582 583 584 585 586 587 588 589 590 591 592

45 1830 1840 1850 1860 1870 1880
 • • • • • •
GGA GCG AAC CTC AAC CTC TCC TGC CAC TCG GCC TCT AAC CCA TCC CCG CAG TAT TCT
Gly Ala Asn Leu Asn Leu Ser Cys His Ser Ala Ser Asn Pro Ser Pro Gln Tyr Ser
574 575 576 577 578 579 580 581 582 583 584 585 586 587 588 589 590 591 592

50 1830 1840 1850 1860 1870 1880
 • • • • • •
GGA GCG AAC CTC AAC CTC TCC TGC CAC TCG GCC TCT AAC CCA TCC CCG CAG TAT TCT
Gly Ala Asn Leu Asn Leu Ser Cys His Ser Ala Ser Asn Pro Ser Pro Gln Tyr Ser
574 575 576 577 578 579 580 581 582 583 584 585 586 587 588 589 590 591 592

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1890 1900 1910 1920 1930
 TGG CGT ATC AAT GGG ATA CCG CAG CAA CAC ACA CAA GTT CTC TTT ATC GCC AAA ATC
 Trp Arg Ile Asn Gly Ile Pro Gln Gln His Thr Gln Val Leu Phe Ile Ala Lys Ile
 593 594 595 596 597 598 599 600 601 602 603 604 605 606 607 608 609 610 611

1940 1950 1960 1970 1980 1990
 ACG CCA AAT AAT AAC GGG ACC TAT GCC TGT TTT GTC TCT AAC TTG GCT ACT GGC CGC
 Thr Pro Asn Asn Asn Gly Thr Tyr Ala Cys Phe Val Ser Asn Leu Ala Thr Gly Arg
 612 613 614 615 616 617 618 619 620 621 622 623 624 625 626 627 628 629 630

2000 2010 2020 2030 2040 2050
 AAT AAT TCC ATA GTC AAG AGC ATC ACA GTC TCT GCA TCT GGA ACT TCT CCT GGT CTC
 Asn Asn Ser Ile Val Lys Ser Ile Thr Val Ser Ala Ser Gly Thr Ser Pro Gly Leu
 631 632 633 634 635 636 637 638 639 640 641 642 643 644 645 646 647 648 649

2060 2070 2080 2090 2100 2110
 TCA GCT GGG GCC ACT GTC GGC ATC ATG ATT GGA GTG CTG GTT GGG GTT GCT CTG ATA
 Ser Ala Gly Ala Thr Val Gly Ile Met Ile Gly Val Leu Val Gly Val Ala Leu Ile
 650 651 652 653 654 655 656 657 658 659 660 661 662 663 664 665 666 667 668

2120 2130 2140 2150 2160
 TAG CAG CCC TGG TGT AGT TTC TTC ATT TCA GGA AGA CTG ACA GTT GTT TTG CTT CTT

2170 2180 2190 2200 2210 2220
 CCT TAA AGC ATT TGC AAC AGC TAC AGT CTA AAA TTG CTT CTT TAC CAA GGA TAT TTA

2230 2240 2250 2260 2270 2280
 CAG AAA ATA CTC TGA CCA GAG ATC GAG ACC ATC CTA GCC AAC ATC GTG AAA CCC CAT

2290 2300 2310 2320 2330
 CTC TAC TAA AAA TAC AAA AAT GAG CTG GGC TTG GTG GCG CGC ACC TGT AGT CCC AGT

2340 2350 2360 2370 2380 2390
 TAC TCG GGA GGC TGA GGC AGG AGA ATC GCT TGA ACC CGG GAG GTG GAG ATT GCA GTG

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2400 2410 2420 2430 2440 2450
 AGC CCA GAT CGC ACC ACT GCA CTC CAG TCT GGC AAC AGA GCA AGA CTC CAT CTC AAA
 5
 2460 2470 2480 2490 2500
 AAG AAA AGA AAA GAA GAC TCT GAC CTG TAC TCT TGA ATA CAA GTT TCT GAT ACC ACT
 10
 2510 2520 2530 2540 2550 2560
 GCA CTG TCT GAG AAT TTC CAA AAC TTT AAT GAA CTA ACT GAC AGC TTC ATG AAA CTG
 15
 2570 2580 2590 2600 2610 2620
 TCC ACC AAG ATC AAG CAG AGA AAA TAA TTA ATT TCA TGG GGA CTA AAT GAA CTA ATG
 20
 2630 2640 2650 2660 2670 2680
 AGG ATA ATA TTT TCA TAA TTT TTT ATT TGA AAT TTT GCT GAT TCT TTA AAT GTC TTG
 25
 2690 2700 2710 2720 2730
 TTT CCC AGA TTT CAG GAA ACT TTT TTT CTT TTA AGC TAT CCA CTC TTA CAG CAA TTT
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 2740 2750 2760 2770 2780 2790
 GAT AAA ATA TAC TTT TGT GAA CAA AAA TTG AGA CAT TTA CAT TTT ATC CCT ATG TGG
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 2800 2810 2820 2830
 TCG CTC CAG ACT TGG GAA ACT ATT CAT GAA TAT TTA TAT TGT ATG
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CEA-(c):

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10 30 50
CAGCCGTGCTCGAAGCGTTCCTGGAGCCCAAGCTCTCCTCCACAGGTGAAGACAGGGCCA

70 90 110
GCAGGAGACACCATGGGGCACCCTCTCAGCCCCACTTCACAGAGTGCGTGTACCCTGGCAG
MetGlyHisLeuSerAlaProLeuHisArgValArgValProTrpGln

130 150 170
GGGCTTCTGCTCACAGCCTCACTTCTAACCTTCTGGAACCCGCCCACCACTGCCCAGCTC
GlyLeuLeuLeuThrAlaSerLeuLeuThrPheTrpAsnProProThrThrAlaGlnLeu

190 210 230
ACTACTGAATCCATGCCATTCAATGTTGCAGAGGGGAAGGAGGTTCTTCTCCTTGTCAC
ThrThrGluSerMetProPheAsnValAlaGluGlyLysGluValLeuLeuLeuValHis

250 270 290
AATCTGCCCCAGCAACTTTTTGGCTACAGCTGGTACAAAGGGGAAAGAGTGGATGGCAAC
AsnLeuProGlnGlnLeuPheGlyTyrSerTrpTyrLysGlyGluArgValAspGlyAsn

310 330 350
CGTCAAATTGTAGGATATGCAATAGGAACCAACAAGCTACCCCAGGGCCCGCAAACAGC
ArgGlnIleValGlyTyrAlaIleGlyThrGlnGlnAlaThrProGlyProAlaAsnSer

370 390 410
GGTCGAGAGACAATATACCCCAATGCATCCCTGCTGATCCAGAACGTCACCCAGAATGAC
GlyArgGluThrIleTyrProAsnAlaSerLeuLeuIleGlnAsnValThrGlnAsnAsp

430 450 470
ACAGGATTCTACACCCTACAAGTCATAAAGTCAGATCTTGTGAATGAAGAAGCAACTGGA
ThrGlyPheTyrThrLeuGlnValIleLysSerAspLeuValAsnGluGluAlaThrGly

490 510 530

CAGTTCCATGTATACCCGGAGCTGCCCAAGCCCTCCATCTCCAGCAACAACCTCCAACCCT
GlnPheHisValTyrProGluLeuProLysProSerIleSerSerAsnAsnSerAsnPro

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550

570

590

GTGGAGGACAAGGATGCTGTGGCCTTCACCTGTGAACCTGAGACTCAGGACACAACCTAC
ValGluAspLysAspAlaValAlaPheThrCysGluProGluThrGlnAspThrThrTyr

10

610

630

650

CTGTGGTGGATAAACAATCAGAGCCTCCCGGTCAGTCCCAGGCTGCAGCTGTCCAATGGC
LeuTrpTrpIleAsnAsnGlnSerLeuProValSerProArgLeuGlnLeuSerAsnGly

15

670

690

710

AACAGGACCCTCACTCTACTCAGTGTCAAGGAATGACACAGGACCCTATGAGTGTGAA
AsnArgThrLeuThrLeuLeuSerValThrArgAsnAspThrGlyProTyrGluCysGlu

20

730

750

770

ATACAGAACCCAGTGAGTGCGAACCGCAGTGACCCAGTCACCTTGAATGTCACCTATGGC
IleGlnAsnProValSerAlaAsnArgSerAspProValThrLeuAsnValThrTyrGly

25

790

810

830

CCGGACACCCCCACCATTTCCCTTCAGACACCTATTACCGTCCAGGGGGCAAACCTCAGC
ProAspThrProThrIleSerProSerAspThrTyrTyrArgProGlyAlaAsnLeuSer

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850

870

890

CTCTCCTGCTATGCAGCCTCTAACCCACCTGCACAGTACTCCTGGCTTATCAATGGAACA
LeuSerCysTyrAlaAlaSerAsnProProAlaGlnTyrSerTrpLeuIleAsnGlyThr

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910

930

950

TTCCAGCAAAGCACACAAGAGCTCTTTATCCCTAACATCACTGTGAATAATAGTGGATCC
PheGlnGlnSerThrGlnGluLeuPheIleProAsnIleThrValAsnAsnSerGlySer

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970

990

1010

TATACCTGCCACGCCAATAACTCAGTCACTGGCTGCAACAGGACCACAGTCAAGACGATC
TyrThrCysHisAlaAsnAsnSerValThrGlyCysAsnArgThrThrValLysThrIle

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ATAGTCACTGAGCTAAGTCCAGTAGTAGCAAAAGCCCCAAATCAAAGCCAGCAAGACCACA
IleValThrGluLeuSerProValValAlaLysProGlnIleLysAlaSerLysThrThr

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1090

1110

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GTCACAGGAGATAAGGACTCTGTGAACCTGACCTGCTCCACAAATGACACTGGAATCTCC
ValThrGlyAspLysAspSerValAsnLeuThrCysSerThrAsnAspThrGlyIleSer

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1150

1170

1190

ATCCGTTGGTTCTTCAAAAACCAGAGTCTCCCGTCCTCGGAGAGGATGAAGCTGTCCCAG
IleArgTrpPhePheLysAsnGlnSerLeuProSerSerGluArgMetLysLeuSerGln

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1210

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GGCAACACCACCCTCAGCATAAACCCTGTCAAGAGGGAGGATGCTGGGACGTATTGGTGT
GlyAsnThrThrLeuSerIleAsnProValLysArgGluAspAlaGlyThrTyrTrpCys

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1270

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1310

GAGGTCTTCAACCCAATCAGTAAGAACCAAAGCGACCCCATCATGCTGAACGTAAACTAT
GluValPheAsnProIleSerLysAsnGlnSerAspProIleMetLeuAsnValAsnTyr

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1330

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AATGCTCTACCACAAGAAAATGGCCTCTCACCTGGGGCCATTGCTGGCATTGTGATTGGA
AsnAlaLeuProGlnGluAsnGlyLeuSerProGlyAlaIleAlaGlyIleValIleGly

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1430

GTAGTGGCCCTGGTTGCTCTGATAGCAGTAGCCCTGGCATGTTTTCTGCATTTCCGGGAAG
ValValAlaLeuValAlaLeuIleAlaValAlaLeuAlaCysPheLeuHisPheGlyLys

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1450

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ACCGGCAGGGCAAGCGACCAGCGTGATCTCACAGAGCACAAACCCTCAGTCTCCAACCAC
ThrGlyArgAlaSerAspGlnArgAspLeuThrGluHisLysProSerValSerAsnHis

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1530

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ACTCAGGACCACTCCAATGACCCACCTAACAAGATGAATGAAGTTACTTATTCTACCCTG
ThrGlnAspHisSerAsnAspProProAsnLysMetAsnGluValThrTyrSerThrLeu

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1570

1590

1610

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AACTTTGAAGCCCAGCAACCCACACAACCAACTTCAGCCTCCCCATCCCTAACAGCCACA
AsnPheGluAlaGlnGlnProThrGlnProThrSerAlaSerProSerLeuThrAlaThr

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1630

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GAAATAATTTATTTCAGAAGTAAAAAAGCAGTAATGAAACCTGTCCTGCTCACTGCAGTGC
GluIleIleTyrSerGluValLysLysGln

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TGATGTATTTCAAGTCTCTCACCCCTCATCACTAGGAGATTCCCTTCCCCTGTAGGGTAGA

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1750

1770

1790

GGGGTGGGGACAGAAACAACTTCTCCTACTCTTCCTTCCTAATAGGCATCTCCAGGCTG

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1810

1830

1850

CCTGGTCACTGCCCCCTCTCTCAGTGTCAATAGATGAAAGTACATTGGGAGTCTGTAGGAA

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1870

1890

1910

ACCCAACCTTCTTGTCATTGAAATTTGGCAAAGCTGACTTTGGGAAAGAGGGACCAGAAC

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1930

1950

1970

TTCCCCTCCCTTCCCCTTTTCCCAACCTGGACTTGTTTTAACTTGCTGTTCAGAGCAC

35

1990

2010

2030

TCATTCTTCCCACCCCCAGTCCTGTCTATCACTCTAATTCCGATTTGCCATAGCCTTG

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2050

2070

2090

AGGTTATGTCCTTTTCCATTAAGTACATGTGCCAGGAAACAGCGAGAGAGAGAAAGTAAA

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2110

2130

2150

CGGCAGTAATGCTTCTCCTATTTCTCCAAAGCCTTGTTGTGAACTAGCAAAGAGAAGAAAA

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2170

2190

2210

TCAAATATATAACCAATAGTGAAATGCCACAGGTTTGTCCACTGTCAGGGTTGTCTACCT

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2230 2250 2270
GTAGGATCAGGGTCTAAGC[.]ACCTTGGTGCTTAGCTAGAATACCACCTAATCCTTCTGGCA[.]
5
2290 2310 2330
AGCCTGTCTTCAGAGAACC[.]CACTAGAAGCA[.]ACTAGGAAAAATCACTTGCCAAAATCCAAG[.]
10
2350 2370 2390
GCAATTCCTGATGGAAAATGCAAAAGCACATATATGTTTTAATATCTTTATGGGCTCTGT[.]
15
2410 2430 2450
TCAAGGCAGTGCTGAGAGGGAGGGGTTATAGCTTCAGGAGGGAACCAGCTTCTGATAAAC[.]
20
2470 2490 2510
ACAATCTGCTAGGA[.]ACTTGGGAAAGGAATCAGAGAGCTGCC[.]CTTCAGCGATTATTTAAAT[.]
25
2530 2550 2570
TGTTAAAGAATACACAATTGGGGTATTGGGATTTTCTCCTTTTCTCTGAGACATTCCA[.]
30
2590 2610 2630
CCATTTTAATTTTTGTA[.]ACTGCTTATTTATGTGAAAAGGGTTATTTTACTTAGCTTAGC[.]
35
2650 2670 2690
TATGTCAGCCAATCCGATTGCCTTAGGTGAAAGAAACCA[.]CCGAAATCCCTCAGGTCCCTT[.]
40
2710 2730 2750
GGTCAGGAGCCTCTCAAGATTTTTTTTGT[.]CAGAGGCTCCAAATAGAAAATAAGAAAAGGT[.]
45
2770 2790 2810
TTTCTTCATT[.]CATGGCTAGAGCTAGATTTAACTCAGTTTCTAGGCACCTCAGACCAATCA[.]
50
2830 2850 2870
TCAACTACCATTCTATTCCATGTTTGCACCTGTGCATTTTCTGTTTGGCC[.]CCCATTCACTT[.]
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2890 2910 2930
 5 TGT CAGGAAACCTTGGCCTCTGCTAAGGTGTATTTGGTCCCTGAGAAGTGGGAGCACCCCT
 2950 2970 2990
 10 ACAGGGACACTATCACTCATGCTGGTGGCATTGTTTACAGCTAGAAAAGCTGCACTGGTGCT
 3010 3030 3050
 15 TAATGCCCCCTTGGGAAATGGGGCTGTGAGGAGGAGGATTATAACTTAGGCCTAGCCTCTT
 3070 3090 3110
 TTAACAGCCTCTGAAATTTATCTTTTCTTCTATGGGGTCTATAAATGTATCTTATAATAA
 20 3130 3150 3170
 AAAGGAAGGACAGGAGGAAGACAGGCAAATGTACTTCTCACCCAGTCTTCTACACAGATG
 25 3190 3210 3230
 GAATCTCTTTGGGGCTAAGAGAAAGGTTTATTCTATATTGCTTACCTGATCTCATGTTA
 30 3250 3270 3290
 GGCTAAGAGGCTTTCTCCAGGAGGATTAGCTTGGAGTTCTCTATACTCAGGTACCTCTT
 35 3310 3330 3350
 TCAGGGTTTTCTAACCCTGACACGGACTGTGCATACTTTCCCTCATCCATGCTGTGCTGT
 40 3370 3390 3410
 GTTATTTAAATTTTCTGGCTAAGATCATGTCTGAATTATGTATGAAAATTATCTATGT
 45 3430 3450
 TTTTATAATAAAAATAATATATCAGACATCGAAAAAAAAA

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(d)

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	580	590	600	610	620	
5	6AG 3AC AAG 6AT GCT 6TG 6CC TCC ACC 16T 6AA CCT 6AG 6TT CAG AAC ACA ACC TAC					
	61u Asp Lys Asp Ala Val Ala Phe Thr Cys 61u Pro 61u Val 61n Asn Thr Thr Tyr					
	124 125 126 127 128 129 130 131 132 133 134 135 136 137 138 139 140 141					
	630	640	650	660	670	680
10	CTG TGG TGG 6TA AAT 66T CAG AGC CTC CCG 6TC AGT CCC AGG CTG CAG CTG TCC AAT					
	Leu Trp Val Asn Gly 61n Ser Leu Pro Val Ser Pro Arg Leu 61n Leu Ser Asn					
	143 144 145 146 147 148 149 150 151 152 153 154 155 156 157 158 159 160					
	690	700	710	720	730	740
15	66C AAC AGG ACC CTC ACT CTA CTC AGC 6TC AAA AGG AAC 6AT 6CA 6GA TCG TAT 6AA					
	Gly Asn Arg Thr Leu Thr Leu Leu Ser Val Lys Arg Asn Asp Ala 61y Ser Tyr 61u					
	161 162 163 164 165 166 167 168 169 170 171 172 173 174 175 176 177 178					
	750	760	770	780	790	800
20	TGT 6AA ATA CAG AAC CCA 6CG AGT 6CC AAC 6CG AGT 6AC CCA 6TC ACC CTG AAT 6TC					
	Cys 61u Ile 61n Asn Pro Ala Ser Ala Asn Arg Ser Asp Pro Val Thr Leu Asn Val					
	181 182 183 184 185 186 187 188 189 190 191 192 193 194 195 196 197 198					
	810	820	830	840	850	
25	CTC TAT GGC CCA 6AT 6GC CCC ACC ATT TCC CCC TCA AAG GCC AAT TAC CGT CCA 666					
	Leu Tyr Gly Pro Asp Gly Pro Thr Ile Ser Pro Ser Lys Ala Asn Tyr Arg Pro Gly					
	201 202 203 204 205 206 207 208 209 210 211 212 213 214 215 216 217 218					
	860	870	880	890	900	910
30	GAA AAT CTG AAC CTC TCC TGC CAC 6CA 6CC TCT AAC CCA CCT 6CA CAG TAC TCT TGG					
	61u Asn Leu Asn Leu Ser Cys His Ala Ala Ser Asn Pro Pro Ala 61n Tyr Ser Trp					
	219 220 221 222 223 224 225 226 227 228 229 230 231 232 233 234 235 236					
	920	930	940	950	960	970
35	TTT ATC AAT 6GG ACB TTC CAG CAA TCC ACA CAA 6AG CTC TTT ATC CCC AAC ATC ACT					
	Phe Ile Asn Gly Thr Phe 61n 61n Ser Thr 61n 61u Leu Phe Ile Pro Asn Ile Thr					
	237 238 239 240 241 242 243 244 245 246 247 248 249 250 251 252 253 254					
	980	990	1000	1010	1020	
40	6TG AAT AAT AGC 6GA TCC TAT ATG TGC CAA 6CC CAT AAC TCA 6CC ACT 66C CTC AAT					
	Val Asn Asn Ser Gly Ser Tyr Met Cys 61n Ala His Asn Ser Ala Thr Gly Leu Asn					
	255 256 257 258 259 260 261 262 263 264 265 266 267 268 269 270 271 272					
	1030	1040	1050	1060	1070	1080
45	AGG ACC ACA 6TC ACB ATG ATC ACA 6TC TCT 6GA AGT 6CT CCT 6TC CTC TCA 6CT 6TG					
	Arg Thr Thr Val Thr Met Ile Thr Val Ser Gly Ser Ala Pro Val Leu Ser Ala Val					
	273 274 275 276 277 278 279 280 281 282 283 284 285 286 287 288 289 290					
	1090	1100	1110	1120	1130	1140
50	6CC ACC 6TC 66C ATC ACB ATT 66A 6TG CTG 6CC AGG 6TG 6CT CTG ATA TAG CAG CCC					
	Ala Thr Val Gly Ile Thr Ile Gly Val Leu Ala Arg Val Ala Leu Ile ---					
	291 292 293 294 295 296 297 298 299 300 301 302 303 304 305 306 307 308					
55						

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1150 1160 1170 1180 1190
TGG TGT ATT TTC GAT ATT TCA GGA AGA CTG GCA GAT TGG ACC AGA CCC TGA ATT CTT

10

1200 1210 1220 1230 1240 1250
CTA GCT CCT CCA ATC CCA TTT TAT CCC ATG GAA CCA CTA AAA ACA AGG TCT GCT CTG

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1260 1270 1280 1290 1300 1310
CTC CTG AAG CCC TAT ATG CTG GAG ATG GAC AAC TCA ATG AAA ATT TAA AGG AAA AAC

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1320 1330 1340 1350 1360 1370
CCT CAG GCC TGA GGT GTG TGC CAC TCA GAG ACT TCA CCT AAC TAG AGA CAG GCA AAC

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1380 1390 1400 1410 1420
TGC AAA CCA nnc CTC TTT CCG TTG GCA GGA TGA TGG TGT CAT TAG TAT TTC ACA AGA

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1430 1440 1450 1460 1470 1480
AGT ACC TTC AGA GGG TAA CTT AAC AGA GTA TCA GAT CTA TCT TGT CAA TCC CAA GGT

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1490 1500 1510 1520 1530 1540
TTT ACA TAA AAT AAG CGA TCC TTT AGT GCA CCC AGT GAC TGA CAT TAG CAG CAT CTT

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1550 1560 1570 1580 1590
TAA CAC AGC CBT GTG TTC AAG TGT ACA GTG GTC CTT TTC AGA GTT GGA nnt ACT CCA

45

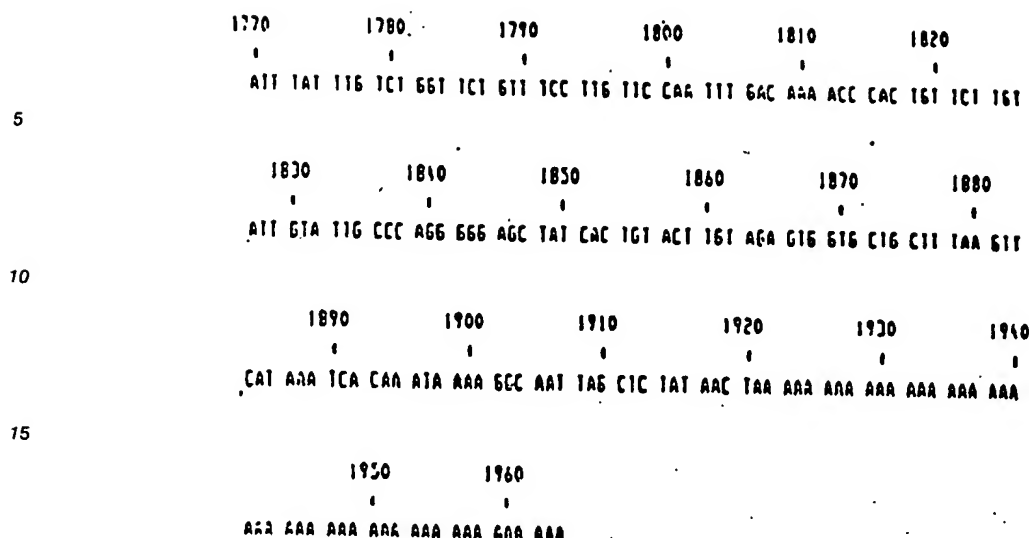
1600 1610 1620 1630 1640 1650
ACT GAA ATG TTA AGG AAG AAG ATA GAT CCA ATT AAA AAA AAT TAA AGC CAA TTT AAA

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1660 1670 1680 1690 1700 1710
AGA AAA AAA GAA CAC AGG AGA TTC CAG TCT ACT TGA GTT AGC ATA ATA CAG AAG TCC

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1720 1730 1740 1750 1760
CCT CTA CTT TAA CTT TTA CAA AAA AGT AAC CTG AAC TAA TCT GAT GTT AAC CAA TGT



A schematic relationship of the transmembrane CEA's, namely TM-1 (CEA-(c)), TM-2 (CEA-(e)), TM-3 (CEA-(f)) and TM-4 (CEA-(g)) is depicted in Fig. 1:

Assuming TM-1 is composed of five sections as depicted in Fig. 1, namely 10, 12, 14, 16 and 18, TM-2 differs from TM-1 in that the 100 amino acid (100 AA) section 14 is deleted and at splice point 20 between sections 12 and 16, surprisingly an extra amino acid, namely Asp occurs.

TM-3 is the same as TM-1 except that section 18 is truncated at splice point 22, i.e., a section of 70 amino acids is deleted and results in a new section made up of subsections 24 + 26. Surprisingly, however, six new amino acids (section 26) occur. Another example of formation of a novel amino acid sequence resulting from a deletion of nucleic acid sequence is for platelet derived growth factor-A.

TM-4 is the same as TM-2 up until the end of subsection 24.

Subsection 24 is contained in section 18 of TM-1 and TM-2, but is not depicted in Fig. 1 for TM-1 and TM-2.

Some CEA epitopes are unique. These are the epitopes which have been useful for distinguishing the various CEA-like antigens immunologically. Peptide epitopes are defined by the linear amino acid sequence of the antigen and/or features resulting from protein folding. The information required for protein folding is encoded in the primary amino acid sequence. Therefore, antigenic differences ultimately result from differences in the primary structure of the different CEA molecules. The differences residing in the CEA protein in the CEA species can thus be determined by determining the primary amino acid sequences. This can be most readily accomplished by cloning and sequencing each of the genes for CEA. To determine which gene products will be most useful for cancer diagnosis, unique probes can be selected for each gene and expression of each gene can be determined in different tumor types by nucleic acid hybridization techniques. The present invention provides a tool with which to identify potential genes coding for different members of the CEA family and to determine the theoretical primary amino acid sequences for them. Using the method of automated peptide synthesis, peptides can then be synthesized corresponding to unique sequences in these antigens. With these peptides, antibodies to these sequences can be produced which, in the intact CEA molecule, might not be recognized by the animal being immunized. Having accomplished this, advantage can then be taken of the differences in these antigens to generate specific immunoassays for the measurement of each antigen.

A wide variety of host/cloning vehicle combinations may be employed in cloning the double-stranded nucleic acid prepared in accordance with this invention. For example, useful cloning vehicles may consist of segments of chromosomal, non-chromosomal and synthetic DNA sequences, such as various known derivatives of SV40 and known bacterial plasmids, e.g., plasmids from *E. coli* including col E1, pCR1, pBR322, pMB89 and their derivatives, wider host range plasmids, e.g., RP4, and phage DNAs, e.g., the numerous derivatives of phage, e.g., NM989, and other DNA phages, e.g., M13 and Filamentous single-stranded DNA phages and vectors derived from combinations of plasmids and phage DNAs such as plasmids which have been modified to employ phage DNA or other expression control sequences or yeast plasmids such as the 2 μ plasmid or derivatives thereof. Useful hosts may include bacterial hosts such as strains of *E. coli*, such as *E. coli* HB 101, *E. coli* X1776, *E. coli* X2282, *E. coli* MRC1 and strains of

Pseudomonas, *Bacillus subtilis*, *Bacillus stearothermophilus* and other *E. coli*, bacilli, yeasts and other fungi, animal or plant hosts such as animal (including human) or plant cells in culture or other hosts. Of course, not all host/vector combinations may be equally efficient. The particular selection of host/cloning vehicle combination may be made by those of skill in the art after due consideration of the principles set forth

without departing from the scope of this invention.

Furthermore, within each specific cloning vehicle, various sites may be selected for insertion of the nucleic acid according to the present invention. These sites are usually designated by the restriction endonuclease which cuts them. For example, in pBR322 the PstI site is located in the gene for beta-lactamase, between the nucleotide triplets that code for amino acids 181 and 182 of that protein. One of the two HindIII endonuclease recognition sites is between the triplets coding for amino acids 101 and 102 and one of the several Taq sites at the triplet coding for amino acid 45 of beta-lactamase in pBR322. In similar fashion, the EcoRI site and the PvuII site in this plasmid lie outside of any coding region, the EcoRI site being located between the genes coding for resistance to tetracycline and ampicillin, respectively. These sites are well recognized by those of skill in the art. It is, of course, to be understood that a cloning vehicle useful in this invention need not have a restriction endonuclease site for insertion of the chosen DNA fragment. Instead, the vehicle could be cut and joined to the fragment by alternative means.

The vector or cloning vehicle and in particular the site chosen therein for attachment of a selected nucleic acid fragment to form a recombinant nucleic acid molecule is determined by a variety of factors, e.g., the number of sites susceptible to a particular restriction enzyme, the size of the protein to be expressed, the susceptibility of the desired protein to proteolytic degradation by host cell enzymes, the contamination of the protein to be expressed by host cell proteins difficult to remove during purification, the expression characteristics, such as the location of start and stop codons relative to the vector sequences, and other factors recognized by those of skill in the art. The choice of a vector and an insertion site for a particular gene is determined by a balance of these factors, not all sections being equally effective for a given case.

Methods of inserting nucleic acid sequences into cloning vehicles to form recombinant nucleic acid molecules include, for example, dA-dT tailing, direct ligation, synthetic linkers, exonuclease and polymerase-linked repair reactions followed by ligation, or extension of the nucleic acid strand with an appropriate polymerase and an appropriate single-stranded template followed by ligation.

It should also be understood that the nucleotide sequences or nucleic acid fragments inserted at the selected site of the cloning vehicle may include nucleotides which are not part of the actual structural gene for the desired polypeptide or mature protein or may include only a fragment of the complete structural gene for the desired protein or mature protein.

The cloning vehicle or vector containing the foreign gene is employed to transform an appropriate host so as to permit that host to replicate the foreign gene and to express the protein coded by the foreign gene or portion thereof. The selection of an appropriate host is also controlled by a number of factors recognized by the art. These include, for example, the compatibility with the chosen vector, the toxicity of proteins encoded by the hybrid plasmid, the ease of recovery of the desired protein, the expression characteristics, biosafety and costs. A balance of these factors must be struck with the understanding that not all hosts may be equally effective for expression of a particular recombinant DNA molecule.

The level of production of a protein is governed by two major factors: the number of copies of its gene within the cell and the efficiency with which those gene copies are transcribed and translated. Efficiency of transcription and translation (which together comprise expression) is in turn dependent upon nucleotide sequences, normally situated ahead of the desired coding sequence. These nucleotide sequences or expression control sequences define inter alia, the location at which RNA polymerase interacts to initiate transcription (the promoter sequence) and at which ribosomes bind and interact with the mRNA (the product of transcription) to initiate translation. Not all such expression control sequences function with equal efficiency. It is thus of advantage to separate the specific coding sequences for the desired protein from their adjacent nucleotide sequences and fuse them instead to other known expression control sequences so as to favor higher levels of expression. This having been achieved, the newly engineered nucleic acid, e.g., DNA, fragment may be inserted into a multicopy plasmid or a bacteriophage derivative in order to increase the number of gene copies within the cell and thereby further improve the yield of expressed protein.

Several expression control sequences may be employed as described above. These include the operator, promoter and ribosome binding and interaction sequences (including sequences such as the Shine-Dalgarno sequences) of the lactose operon of *E. coli* ("the lac system"), the corresponding sequences of the tryptophan synthetase system of *E. coli* ("the trp system"), the major operator and promoter regions of phage λ ($O_L P_L$ and $O_R P_R$), the control region of Filamentous single-stranded DNA phages, or other sequences which control the expression of genes of prokaryotic or eukaryotic cells and

their viruses. Therefore, to improve the production of a particular polypeptide in an appropriate host, the gene coding for that polypeptide may be selected and removed from a recombinant nucleic acid molecule containing it and reinserted into a recombinant nucleic acid molecule closer or in a more appropriate relationship to its former expression control sequence or under the control of one of the above described expression control sequences. Such methods are known in the art.

As used herein "relationship" may encompass many factors, e.g., the distance separating the expression enhancing and promoting regions of the recombinant nucleic acid molecule and the inserted nucleic acid sequence, the transcription and translation characteristics of the inserted nucleic acid sequence or other sequences in the vector itself, the particular nucleotide sequence of the inserted nucleic acid sequence and other sequences of the vector and the particular characteristics of the expression enhancing and promoting regions of the vector.

Further increases in the cellular yield of the desired products depend upon an increase in the number of genes that can be utilized in the cell. This is achieved, for illustration purposes, by insertion of recombinant nucleic acid molecules engineered into the temperate bacteriophage λ (NM989), most simply by digestion of the plasmid with a restriction enzyme, to give a linear molecule which is then mixed with a restricted phage λ cloning vehicle (e.g., of the type described by N. E. Murray et al, "Lambdoid Phages That Simplify the Recovery of In Vitro Recombinants", *Molec. Gen. Genet.*, 150, pp. 53-61 (1977) and N. E. Murray et al, "Molecular Cloning of the DNA Ligase Gene From Bacteriophage T4", *J. Mol. Biol.*, 132, pp. 493-505 (1979)) and the recombinant DNA molecule recircularized by incubation with DNA ligase. The desired recombinant phage is then selected as before and used to lysogenize a host strain of *E. coli*.

Particularly useful λ cloning vehicles contain a temperature-sensitive mutation in the repression gene *cl* and suppressible mutations in gene *S*, the product of which is necessary for lysis of the host cell, and gene *E*, the product of which is major capsid protein of the virus. With this system, the lysogenic cells are grown at 32°C and then heated to 45°C to induce excision of the prophage. Prolonged growth at 37°C leads to high levels of production of the protein, which is retained within the cells, since these are not lysed by phage gene products in the normal way, and since the phage gene insert is not encapsulated it remains available for further transcription. Artificial lysis of the cells then releases the desired product in high yield.

In addition, it should be understood that the yield of polypeptides prepared in accordance with this invention may also be improved by substituting different codons for some or all of the codons of the present DNA sequences, these substituted codons coding for amino acids identical to those coded for by the codons replaced.

Finally, the activity of the polypeptides produced by the recombinant nucleic acid molecules of this invention may be improved by fragmenting, modifying or derivatizing the nucleic acid sequences or polypeptides of this invention by well-known means, without departing from the scope of this invention.

The polypeptides of the present invention include the following:

- (1) the polypeptides expressed by the above described cells,
- (2) polypeptides prepared by synthetic means,
- (3) fragments of polypeptides (1) or (2) above, such fragments produced by synthesis of amino acids or by digestion or cleavage.

Regarding the synthetic peptides according to the invention, chemical synthesis of peptides is described in the following publications: S.B.H. Kent, *Biomedical Polymers*, eds. Goldberg, E.P. and Nakajima, A. (Academic Press, New York), 213-242, (1980); A.R. Mitchell, S.B.H. Kent, M. Engelhard and R.B. Merrifield, *J. Org. Chem.*, 43, 2845-2852, (1978); J.P. Tam, T.-W. Wong, M. Riemen, F.-S. Tjoeng and R.B. Merrifield, *Tet. Letters*, 4033-4036, (1979); S. Mojsov, A.R. Mitchell and R.B. Merrifield, *J. Org. Chem.*, 45, 555-560, (1980); J.P. Tam, R.D. DiMarchi and R.B. Merrifield, *Tet. Letters*, 2851-2854, (1981); and S.B.H. Kent, M. Riemen, M. Le Doux and R.B. Merrifield, *Proceedings of the IV International Symposium on Methods of Protein Sequence Analysis*, (Brookhaven Press, Brookhaven, NY), in press, 1981.

In the Merrifield solid phase procedure, the appropriate sequence of L-amino acids is built up from the carboxyl terminal amino acid to the amino terminal amino acid. Starting with the appropriate carboxyl terminal amino acid attached to a polystyrene (or other appropriate) resin via chemical linkage to a chloromethyl group, benzhydrylamine group, or other reactive group of the resin, amino acids are added one by one using the following procedure. The peptide-resin is:

- (a) washed with methylene chloride;
- (b) neutralized by making for 10 minutes at room temperature with 5% (v/v) diisopropylethylamine (or other hindered base) in methylene chloride;
- (c) washed with methylene chloride;
- (d) an amount of amino acid equal to six times the molar amount of the growing peptide chain is activated by combining it with one-half as many moles of a carbodiimide (e.g., dicyclohexylcarbodiimide,

or diisopropylcarbodiimide) for ten minutes at 0 °C, to form the symmetric anhydride of the amino acid. The amino acid used should be provided originally as the N-alpha-tert-butyloxycarbonyl derivative, with side chains protected with benzyl esters (e.g., aspartic or glutamic acids), benzyl ethers (e.g., serine, threonine, cysteine or tyrosine), benzyloxycarbonyl groups (e.g., lysine) or other protecting groups

commonly used in peptide synthesis;

(e) the activated amino acid is reacted with the peptide-resin for two hours at room temperature, resulting in addition of the new amino acid to the end of the growing peptide chain;

(f) the peptide-resin is washed with methylene chloride;

(g) the N-alpha-(tert-butyloxycarbonyl) group is removed from the most recently added amino acid by reacting with 30 to 65%, preferably 50% (v/v) trifluoroacetic acid in methylene chloride for 10 to 30 minutes at room temperature;

(h) the peptide-resin is washed with methylene chloride;

(i) steps (a) through (h) are repeated until the required peptide sequence has been constructed.

The peptide is then removed from the resin and simultaneously the side-chain protecting groups are removed, by reaction with anhydrous hydrofluoric acid containing 10% v/v of anisole or other suitable (aromatic) scavenger. Subsequently, the peptide can be purified by gel filtration, ion exchange, high pressure liquid chromatography, or other suitable means.

In some cases, chemical synthesis can be carried out without the solid phase resin, in which case the synthetic reactions are performed entirely in solution. The reactions are similar and well known in the art, and the final product is essentially identical.

Digestion of the polypeptide can be accomplished by using proteolytic enzymes, especially those enzymes whose substrate specificity results in cleavage of the polypeptide at sites immediately adjacent to the desired sequence of amino acids.

Cleavage of the polypeptide can be accomplished by chemical means. Particular bonds between amino acids can be cleaved by reaction with specific reagents. Examples include the following: bonds involving methionine are cleaved by cyanogen bromide; asparaginyl-glycine bonds are cleaved by hydroxylamine.

The present invention has the following advantages:

(1) The nucleic acids coding for TM-1, TM-2 and TM-3 can be used as probes to isolate other members of the CEA gene family.

(2) The nucleic acids coding for TM-1, TM-2 and TM-3 can be used to derive oligonucleotide probes to determine the expression of TM-1, TM-2, TM-3 and other CEA genes in various tumor types.

(3) TM-1, TM-2, TM-3 and TM-4 nucleotide sequences can be used to predict the primary amino acid sequence of the protein for production of synthetic peptides.

(4) Synthetic peptides derived from the above sequences can be used to produce sequence-specific antibodies.

(5) Immunoassays for each member of the CEA antigen family can be produced with these sequence-specific antibodies and synthetic peptides.

(6) The aforementioned immunoassays can be used as diagnostics for different types of cancer if it is determined that different members of the CEA family are clinically useful markers for different types of cancer.

Peptides according to the present invention can be labelled by conventional means using radioactive moieties (e.g., ¹²⁵I), enzymes, dyed and fluorescent compounds, just to name a few.

Several possible configurations for immunoassays according to the present invention can be used. The readout systems capable of being employed in these assays are numerous and non-limiting examples of such systems include fluorescent and colorimetric enzyme systems, radioisotopic labelling and detection and chemiluminescent systems. Two examples of immunoassay methods are as follows:

(1) An enzyme linked immunoassay (ELISA) using an antibody preparation according to the present invention (including Fab or F(ab)' fragments derived therefrom) to a solid phase (such as a microtiter plate or latex beads) is attached a purified antibody of a specificity other than that which is conjugated to the enzyme. This solid phase antibody is contacted with the sample containing CEA antigen family members. After washing, the solid phase antibody-antigen complex is contacted with the conjugated anti-peptide antibody (or conjugated fragment). After washing away unbound conjugate, color or fluorescence is developed by adding a chromogenic or fluorogenic substrate for the enzyme. The amount of color or fluorescence developed is proportional to the amount of antigen in the sample.

(2) A competitive fluorometric immunoassay using fluorescently labelled peptide or synthetic peptides of the sequences for TM-2, TM-2, TM-3 and TM-4. In this example, the purified peptide expressed by cells or synthetic peptides thereof are fluorescently labelled. To a solid phase is attached a purified antibody. This solid phase is then contacted with sample containing CEA antigen family members to which has

been added fluorescent peptide probe. After binding, excess probe is washed away the amount of bound probe is quantitated. The amount of bound fluorescent probe will be inversely proportional to the amount of antigen in the sample.

In the nucleic acid hybridization method according to the present invention, the nucleic acid probe is conjugated with a label, for example, an enzyme, a fluorophore, a radioisotope, a chemiluminescent compound, etc. In the most general case, the probe would be contacted with the sample and the presence of any hybridizable nucleic acid sequence would be detected by developing in the presence of a chromogenic enzyme substrate, detection of the fluorophore by epifluorescence, by autoradiography of the radioisotopically labelled probe or by chemiluminescence. The detection of hybridizable RNA sequences can be accomplished by (1) a dot blot methodology or (2) an *in situ* hybridization methodology. Methods for these last two techniques are described by D. Gillespie and J. Bresser, "mRNA Immobilization in Nal: Quick Blots", *Biotechniques*, 184-192, November/December 1983 and J. Lawrence and R. Singer, "Intracellular Localization of Messenger RNAs for Cytoskeletal Proteins", *Cell*, 45, 407-415, May 9, 1986, respectively. The readout systems can be the same as described above, e.g., enzyme labelling, radiolabelling, etc.

As stated above, the invention also relates to the use in medicine of the aforementioned complex of the invention.

The invention further provides a pharmaceutical composition containing as an active ingredient a complex of the invention in the form of a sterile and/or physiologically isotonic aqueous solution.

For parenteral administration, solutions and emulsions containing as an active ingredient the complex of the invention should be sterile and, if appropriate, blood-isotonic.

It is envisaged that the active complex will be administered perorally, parenterally (for example, intramuscularly, intraperitoneally, or intravenously), rectally or locally.

Example 1: Preparation of cDNA in pcE22 which codes for TM2-CEA [CEA-(e)]

Example 1a: RNA Preparation

Messenger RNA was prepared by the proteinase K extraction method of J. Favolaro, R. Treisman and R. Kamen, *Methods in Enzymology*, 65, 718, Academic Press, Inc. (1980), followed by oligo dT cellulose chromatography to yield poly A+ RNA (3'-polyadenylated eukaryotic RNA containing most mRNA sequences that can be translated into polypeptides). To obtain approximately 100 µg of poly A+ RNA, approximately 3×10^8 cells of transfectant 23.411 (ATCC No. CRL 9731, deposited with the ATCC on June 1, 1988), that expresses TM-1, TM-2, TM-3 and TM-4, Kamarck et al, *Proc. Natl. Acad. Sci., USA*, 84, 5350-5354, August 1987, were harvested from roller bottles after late logarithmic growth. Cells were lysed by homogenization in an ice-cold solution of 140 mM NaCl, 1.5 mM MgCl₂, 10 mM Tris-HCl, pH 8.0, 0.5% NP40®, 4 mM dithiothreitol and 20 units of placental ribonuclease inhibitor/ml. Sodium deoxycholate was then added to 0.2%. Cytoplasm and nuclei were separated by centrifugation of the homogenate at 12,000xg for 20 minutes. The cytoplasmic fraction was mixed with an equal volume of 0.2 M Tris-HCl, pH 7.8, 25 mM EDTA, 0.3 M NaCl, 2% sodium dodecyl sulfate and 400 µg/ml of proteinase K, incubated for 1 hour at 37°C, then extracted once with an equal volume of phenol/chloroform (1:1/v/v) solution. Nucleic acids were obtained by ethanol precipitation of the separated aqueous phase. Total RNA was enriched by passage in 0.5 M NaCl, 10 mM Tris-HCl, pH 7.8, 0.1% sarcosyl® through an oligo dT(12-18) cellulose column. After washing, bound RNA was eluted in the same solution without sodium chloride.

Example 1b: Reverse Transcription of mRNA

Ten micrograms of poly A+ RNA were primed for reverse transcription with oligo dT(12-18) and pdN₆ primers. One hundred microliter reaction was performed for 4 hours at 42°C with 200 units AMV reverse transcriptase (Life Science, Inc. St. Petersburg, Florida, U.S.A.). The RNA component of the cDNA/mRNA hybrids was replaced with the second complementary strand by treatment with RNase H, *E. coli* DNA polymerase I and *E. coli* DNA ligase at 12°C and 22°C for 1.5 hours each. Molecular ends were polished by treatment with T₄ DNA polymerase. cDNA was phenol/chloroform extracted and purified over a "SEPHADEX® G-50" spun column prepared in 10 mM Tris-HCl, pH 7.8, 1 mM EDTA (TE).

Example 1c: Cloning of pcE22 (plasmid cDNA E22)

Synthetic DNA linkers	5' pCCCGGG 3'
	3' GGGCCCTTAA 5'

were attached to the ends of cDNA by blunt end ligation with excess T4 DNA ligase. Excess linkers were removed by chromatography through "SEPHADEX® G-50" (medium) in TE, and by fractionation on 0.8% low melting agarose gel. Based on Northern blot analysis of poly A+ RNA of the 23.411 cell line, the size of the CEA-related mRNA was estimated at 3.6 kb. Therefore, cDNA fragments between 2 and 4 kb were recovered from gel slices and fragments were ethanol precipitated. After resuspension of cDNA in TE, EcoRI-cleaved lambda gt10 arms were added to cDNA at an estimated molar ratio of 1:1. Ligation proceeded at 7 °C for 2 days in the presence of T4 DNA ligase. Aliquots of the ligation reaction were added to commercially-obtained packaging mix (Stratagene, San Diego, California, U.S.A.). Five million phage particles were obtained after in vitro packaging and infection of E. coli host NM514.

Example 1d: Screening of Recombinant Library

Five hundred thousand packaged lambda particles were plated on lawns of E. coli NM514 and replicate patterns were lifted onto nitrocellulose sheets as described by W.D. Benton and R.W. Davis, Science 196, 180-182, (1977). Positive phage were selected by hybridization with ³²P-labeled LV7 cDNA insert probe that contained a domain repeated among various CEA family members. By multiple rounds of screening. Phage from individual plaques were amplified and titered, and these were used to prepare small quantities of recombinant phage DNA.

Example 1e: DNA Manipulation

Phage DNA was prepared according to T. Maniatis, E. Fritsch and J. Sambrook, Molecular Cloning, A Laboratory Manual, Cold Spring Harbor, (1982). DNA segments were isolated from low melting agarose gels and inserted for subcloning into Bluescript plasmid vectors (Stratagene, San Diego, California, U.S.A.). DNA sequencing was performed by the dideoxy termination method of F. Sanger, S. Nicklen and A. Coulson, Proc. Natl. Acad. Sci., U.S.A., 74, 5463-5467, (1977). The nucleic acid and translated sequence for cDNA in pcE22 is given hereinabove (TM-2 (CEA-(e))).

Example 2: Preparation of cDNA in pcHT-6 which Partically Codes for TM3-CEA [CEA-(f)]

Example 2a: RNA Preparation

Messenger RNA was prepared by the proteinase K extraction method of J. Favolaro, R. Treisman and R. Kamen, Methods in Enzymology, 65 718, Academic Press, Inc. (1980), followed by oligo dT cellulose chromatography to yield poly A+ RNA (3'-polyadenylated eukaryotic RNA containing most mRNA sequences that can be translated into polypeptides). To obtain approximately 100 ug of poly A+ RNA, approximately 3 x 10⁸ cells of HT-29 tumor cells (ATCC HTB38) were harvested from roller bottles after late logarithmic growth. Cells were lysed by homogenization in an ice-cold solution of 140 mM NaCl, 1.5 mM MgCl₂, 10 mM Tris-HCl, pH 8.0, 0.5% NP40®, 4 mM dithiothreitol and 20 units of placental ribonuclease inhibitor/ml. Sodium deoxycholate was then added to 0.2%. Cytoplasm and nuclei were separated by centrifugation of the homogenate at 12,000xg for 20 minutes. The cytoplasmic fraction was mixed with an equal volume of 0.2 M Tris-HCl, pH 7.8, 25 mM EDTA, 0.3 M NaCl, 2% sodium dodecyl sulfate and 400 µg/ml of proteinase K, incubated for 1 hour at 37 °C, then extracted once with an equal volume of phenol/chloroform (1:1/v:v) solution. Nucleic acids were obtained by ethanol precipitation of the separated aqueous phase. Total RNA was enriched by passage in 0.5 M NaCl, 10 mM Tris-HCl, pH 7.8, 0.1% sarcosyl® through an oligo dT(12-18) cellulose column. After washing, bound RNA was eluted in the same solution without sodium chloride.

Example 2b: Reverse Transcription of mRNA

Ten micrograms of HT-29 poly A+ RNA were primed for reverse transcription with oligo dT(12-18) and pdN₆ primers. One hundred microliter reaction was performed for 4 hours at 42 °C with 200 units AMV reverse transcriptase (Life Science, Inc. St. Petersburg, Florida, U.S.A.). The RNA component of the cDNA/mRNA hybrids was replaced with the second complementary strand by treatment with RNase H, E. coli DNA polymerase I and E. coli DNA ligase at 12 °C and 22 °C for 1.5 hours each. Molecular ends were polished by treatment with T4 DNA polymerase. cDNA was phenol/chloroform extracted and purified over a "SEPHADEX® G-50" spun column prepared in 10 mM Tris-HCl, pH 7.8, 1 mM EDTA (TE).

Example 2c: Cloning of pcHT-6 (plasmid cDNA HT-6)

Synthetic DNA linkers 5' pCCCGGG 3'
 3' GGGCCCTAA 5'

- 5 were attached to the ends of cDNA by blunt end ligation with excess T4 DNA ligase. Excess linkers were removed by chromatography through "SEPHADEX® G-50" (medium) in TE, and by fractionation on 0.8% low melting agarose gel. Based on Northern blot analysis of poly A+ RNA of the HT-29 cell line, the size of the CEA-related mRNA was estimated at 2.2 kb. Therefore, cDNA fragments between 2 and 3 kb were recovered from gel slices and fragments were ethanol precipitated. After resuspension of cDNA in TE,
- 10 EcoRI-cleaved lambda gt10 arms were added to cDNA at an estimated molar ratio of 1:1. Ligation proceeded at 7°C for 2 days in the presence of T4 DNA ligase. Aliquots of the ligation reaction were added to commercially-obtained packaging mix (Stratagene, San Diego, California, U.S.A.). Five million phage particles were obtained after in vitro packaging and infection of E. coli host NM514.

15 Example 2d: Screening of Recombinant Library

- Five hundred thousand packaged lambda particles were plated on lawns of E. coli NM514 and replicate patterns were lifted onto nitrocellulose sheets as described by W.D. Benton and R.W. Davis, Science, 196, 180-182, (1977). Positive phage were selected by hybridization with ³²P-labeled LV7 cDNA insert probe that
- 20 contained a domain repeated among various CEA family members. By multiple rounds of screening. Phage from individual plaques were amplified and titered, and these were used to prepare small quantities of recombinant phage DNA.

Example 2e: DNA Manipulation

- 25 Phage DNA was prepared according to T. Maniatis, E. Fritsch and J. Sambrook, Molecular Cloning, A Laboratory Manual, Cold Spring Harbor, (1982). DNA segments were isolated from low melting agarose gels and inserted for subcloning into Bluescript plasmid vectors (Stratagene, San Diego, California, U.S.A.). DNA sequencing was performed by the dideoxy termination method of F. Sanger, S. Nicklen and A. Coulson, Proc. Natl. Acad. Sci., U.S.A., 74, 5463-5467, (1977). The nucleic acid and translated sequence for cDNA in
- 30 HT-6 not complete at the 5' end of its coding region, but the nucleotide sequence and restriction map of the HT-6 insert indicates that it is related to nucleic acid sequences of cDNA clones coding for CEA-(c) and CEA-(e). The nucleotide sequence of HT-6 insert differs from these clones at only nucleotide position 1463 to 1515 and 1676 to 2429 of the CEA-(c) cDNA. It is inferred from this result that the pcHT-6 insert is a
- 35 partial coding sequence for CEA-(f) and the presumed nucleic acid and translated sequence of CEA-(f) is given hereinabove (TM-3 (CEA-(f))).

Example 3: Preparation of cDNA which codes for TM4-CEA [CEA-(g)]40 Example 3a: RNA Preparation

- Messenger RNA is prepared by the proteinase K extraction method of J. Favolaro, R. Treisman and R. Kamen, Methods in Enzymology, 65, 718, Academic Press, Inc. (1980), followed by oligo dT cellulose chromatography to yield poly A+ RNA (3'-polyadenylated eukaryotic RNA containing most mRNA sequences that can be translated into polypeptides). To obtain approximately 100 ug of poly A+ RNA,
- 45 approximately 3 x 10⁸ cells of transfectant 23.411 or tumor cell line HT-29 (ATCC HTB 38) are harvested from roller bottles after late logarithmic growth. Cells are lysed by homogenization in an ice-cold solution of 140 mM NaCl, 1.5 mM MgCl₂, 10 mM Tris-HCl, pH 8.0, 0.5% NP40®, 4 mM dithiothreitol and 20 units of placental ribonuclease inhibitor/ml. Sodium deoxycholate was then added to 0.2%. Cytoplasm and nuclei
- 50 are separated by centrifugation of the homogenate at 12,000xg for 20 minutes. The cytoplasmic fraction is mixed with an equal volume of 0.2 M Tris-HCl, pH 7.8, 25 mM EDTA, 0.3 M NaCl, 2% sodium dodecyl sulfate and 400 µg/ml of proteinase K, incubated for 1 hour at 37°C, then extracted once with an equal volume of phenol/chloroform (1:1/v/v) solution. Nucleic acids are obtained by ethanol precipitation of the separated aqueous phase. Total RNA is enriched by passage in 0.5 M NaCl, 10 mM Tris-HCl, pH 7.8, 0.1%
- 55 sarcosyl through an oligo dT(12-18) cellulose column. After washing, bound RNA is eluted in the same solution without sodium chloride.

Example 3b: Reverse Transcription of mRNA

Ten micrograms of 23.411 or HT 29 poly A+ RNA are primed for reverse transcription with oligo dT(12-18) and pdN₆ primers. One hundred microliter reaction was performed for 4 hours at 42°C with 200 units AMV reverse transcriptase (Life Science, Inc. St. Petersburg, Florida, U.S.A.). The RNA component of the cDNA/mRNA hybrids is replaced with the second complementary strand by treatment with RNase H, *E. coli* DNA polymerase I and *E. coli* DNA ligase at 12°C and 22°C for 1.5 hours each. Molecular ends are polished by treatment with T4 DNA polymerase. cDNA is phenol/chloroform extracted and purified over a "SEPHADEX® G-50" spun column prepared in 10 mM Tris-HCl, pH 7.8, 1 mM EDTA (TE).

Example 3c: Cloning of cDNA for CEA-(g)

Synthetic DNA linkers 5' pCCCGGG 3'
 3' GGGCCCTTAA 5'

are attached to the ends of cDNA by blunt end ligation with excess T4 DNA ligase. Excess linkers are removed by chromatography through "SEPHADEX® G-50" (medium) in TE, and by fractionation on 0.8% low melting agarose gel. Based on Northern blot analysis of poly A+ RNA of the 23.411 and HT-29 cell lines, the size of the CEA-related mRNA is estimated at 1.7 kb. Therefore, cDNA fragments between 1 and 2 kb are recovered from gel slices and fragments are ethanol precipitated. After resuspension of cDNA in TE, EcoRI-cleaved lambda gt10 arms are added to cDNA at an estimated molar ratio of 1:1. Ligation proceeds at 7°C for 2 days in the presence of T4 DNA ligase. Aliquots of the ligation reaction are added to commercially-obtained packaging mix (Stratagene, San Diego, California, U.S.A.). Phage particles are obtained after *in vitro* packaging and infection of *E. coli* host NM514.

Example 3d: Screening of Recombinant Library

Five hundred thousand to one million packaged lambda particles are plated on lawns of *E. coli* NM514 and replicate patterns are lifted onto nitrocellulose sheets as described by W.D. Benton and R.W. Davis, Science, 196, 180-182, (1977). Positive phage are selected by hybridization with ³²P-labeled LV7 cDNA insert probe that contained a domain repeated among various CEA family members. By this selection method, positive phage are obtained after multiple rounds of screening. Phage from individual plaques are amplified and titered, and these are used to prepare small quantities of recombinant phage DNA.

Example 3e: DNA Manipulation

Phage DNA is prepared according to T. Maniatis, E. Fritsch and J. Sambrook, Molecular Cloning, A Laboratory Manual, Cold Spring Harbor, (1982). DNA segments are isolated from low melting agarose gels and inserted for subcloning into Bluescript plasmid vectors (Stratagene, San Diego, California, U.S.A.). DNA sequencing is performed by the dideoxy termination method of F. Sanger, S. Nicklen and A. Coulson, Proc. Natl. Acad. Sci., U.S.A., 74, 5463-5467, (1977). The nucleotide and translated sequence for a cDNA coding for CEA-(g) is given hereinabove (TM-4 (CEA-(g))).

Example 4: Screening of a KG-1 cDNA Library with ³²P-labelled CEA Probe, LV7 (CEA-(A))

A segment of cDNA coding for a portion of carcinoembryonic antigen [LV7 or CEA-(a)] was radiolabelled by random priming and used to detect homologous sequences on filter replicas of a commercial cDNA library prepared from KG-1 cells in bacteriophage vector λ gt11 (Clontech Laboratories, Inc., Palo Alto, CA., U.S.A.). Hybridizations were performed at 68°C in 2xSSSPE (1xSSPE - 0.15 M NaCl, 0.01 M sodium phosphate and 1 mM EDTA, pH 7), 5x Denhardt's solution and 100 µg of denatured salmon sperm DNA per ml, and post-hybridization washes were in 0.2xSSC, 0.25% sodium dodecyl sulfate.

Positive phage were picked, rescreened to homogeneity, and amplified for production of DNA. cDNA inserts were excised from phage DNA with EcoRI endonuclease and subcloned into the EcoRI site of the plasmid vector pBluescript KS. DNA sequencing on double-stranded DNA was by the method of Sanger et al, supra. The sequences of two different inserts from the KG-1 cDNA library are shown below:

pcRGCEAl:

	1	acagcacagctgacagccgtactcaggaagcttctggatcctaggcttatctccacagag	60
5	61	gagaacacacaagcagcagagaccatggggccctctcagccctccctgcacacacctc MetGlyProLeuSerAlaProProCysThrHisLeu	120
	121	atcacttggaaggggtcctgctcacagcatcacttttaaacttctggaatccgcccaca IleThrTrpLysGlyValLeuLeuThrAlaSerLeuLeuAsnPheTrpAsnProProThr	180
10	181	actgcccaagtcacgattgaagcccagccacccaaagtcttctgaggggaaggatgttctt ThrAlaGlnValThrIleGluAlaGlnProProLysValSerGluGlyLysAspValLeu	240
	241	ctacttggtccacaatttgccccagaatcttgctggctacatttggtacaaagggcaaagt LeuLeuValHisAsnLeuProGlnAsnLeuAlaGlyTyrIleTrpTyrLysGlyGlnMet	300
15	301	acatacgtctaccattacattacatcatatgtagtagacggtcaaagaattatatatggg ThrTyrValTyrHisTyrIleThrSerTyrValValAspGlyGlnArgIleIleTyrGly	360
	361	cctgcatacagtggaagagaaagagatatattccaatgcatccctgctgatccagaatgtc ProAlaTyrSerGlyArgGluArgValTyrSerAsnAlaSerLeuLeuIleGlnAsnVal	420
20	421	acgcaggaggatgcaggatcctacaccttacacatcataaagcgacgcgatgggactgga ThrGlnGluAspAlaGlySerTyrThrLeuHisIleIleLysArgArgAspGlyThrGly	480
	481	ggagtaactggacatttcaccttcaccttacacctggagactcccaagccctccatctcc GlyValThrGlyHisPheThrPheThrLeuHisLeuGluThrProLysProSerIleSer	540
25	541	agcagcaacttaaatcccagggaggccatggaggctgtgatcttaacctgtgatcctgcg SerSerAsnLeuAsnProArgGluAlaMetGluAlaValIleLeuThrCysAspProAla	600
	601	actccagccgcaagctaccagtggtggatgaatggctcagagcctccctatgactcacagg ThrProAlaAlaSerTyrGlnTrpTrpMetAsnGlyGlnSerLeuProMetThrHisArg	660
30	661	ttgcagctgtccaaaaccaacaggaccctcttttatatttggtgtcaciaagtatattgca LeuGlnLeuSerLysThrAsnArgThrLeuPheIlePheGlyValThrLysTyrIleAla	720
	721	ggaccctatgaatgtgaaatacgaacccagtgagtgccagccgcagtgaccagtcacc GlyProTyrGluCysGluIleArgAsnProValSerAlaSerArgSerAspProValThr	780
35	781	ctgaatctcctcccaaagctgtccaagccctacatcacaaatcaacaacttaaacccccaga LeuAsnLeuLeuProLysLeuSerLysProTyrIleThrIleAsnAsnLeuAsnProArg	840
	841	gagaataaggatgtcttaaccttcacctgtgaacctaaagagtgagaactacacctacatt GluAsnLysAspValLeuThrPheThrCysGluProLysSerGluAsnTyrThrTyrIle	900
40	901	tggtggctaaatgggtcagagcctccctgtcagtcacagggtaaagcgaccattgaaaac TrpTrpLeuAsnGlyGlnSerLeuProValSerProArgValLysArgProIleGluAsn	960
	961	aggatcctcattctacccaatgtcacgagaaatgaaacaggaccttatcaatgtgaaata ArgIleLeuIleLeuProAsnValThrArgAsnGluThrGlyProTyrGlnCysGluIle	1020
45	1021	cgggaccgatattggtggcatccgcagtgaccagtcaccctgaatgtcctctatgggtcca ArgAspArgTyrGlyGlyIleArgSerAspProValThrLeuAsnValLeuTyrGlyPro	1080

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1081	gacctccccagcattttacccttcattcacctattaccggttcaggagaaaacctctacttt	1140
	AspLeuProSerIleTyrProSerPheThrTyrTyrArgSerGlyGluAsnLeuTyrPhe	
1141	tcctgcttcggtgagtcctaaccacgggcacaatatcttggacaattaatgggaagttt	1200
5	SerCysPheGlyGluSerAsnProArgAlaGlnTyrSerTrpThrIleAsnGlyLysPhe	
1201	cagctatcaggacaaaaagctctctatcccccaaataactacaaagcatagtggtctctat	1260
	GlnLeuSerGlyGlnLysLeuSerIleProGlnIleThrThrLysHisSerGlyLeuTyr	
1261	gcttgctctgttcgtaactcagccactggcaaggaaagctccaaatccatcacagtcaaa	1320
10	AlaCysSerValArgAsnSerAlaThrGlyLysGluSerSerLysSerIleThrValLys	
1321	gtctctgactggatattaccctgaattctactagttcctccaattccattttctcccatg	1380
	ValSerAspTrpIleLeuProEnd	
1441	gaatcacgaagagcaagaccactctgttccagaagccctataatctggaggtggacaac	1440
15	tcgatgtaaatttcattgggaaaacccttgctacctgacatgtgagccactcagaactcacc	1500
1441	aaaatggttcgacaccataacaacagctactcaactgtaaaccaggataagaagttgatg	1560
1501	acttcacactgtggacagtttttcaaagatgtcataacaagactcccatcatgacaagg	1620
1561	ctccaccctctactgtctgtctatgcctgcctctttcacttggcaggataatgcagtcac	1680
1621	tagaatttcacatgtagtagcttctgagggtaacaacagagtgctcagatatgtcatctca	1740
1681	acctcaaaacttttacgtaacatctcagggaaaatgtggctctctccatcttgcatacagg	1800
1741	ctcccaatagaaatgaacacagagatattgcctgtgtgtttgcagagaagatgggtttcta	1860
20	1801 taaagagttaggaaagctgaaattatagtagagtctcctttaaatgcacattgtgtggatg	1920
1861	gctctcaccatttcctaagagatacagtgtaaaagacgtgacagtaatactgattctagca	1980
1921	gaataaacatgtaccacatttgcaaaaaa	2010
1981		
25	pcKGEA2:	
1	gggtggatccttaggctcatctccataggggagaaacacacatacagcagagaccatggga	59
	MetGly	
60	cccctctcagccccctccctgcactcagcacatcacctggaaggggctcctgctcacagca	119
30	ProLeuSerAlaProProCysThrGlnHisIleThrTrpLysGlyLeuLeuLeuThrAla	
120	tcactttttaaacttctggaacctgcccaccactgcccagtaataattgaagcccagcca	179
	SerLeuLeuAsnPheTrpAsnLeuProThrThrAlaGlnValIleIleGluAlaGlnPro	
180	cccaaagtttctgaggggaaggatgttcttctacttgtccacaatttgccccagaatctt	239
35	ProLysValSerGluGlyLysAspValLeuLeuLeuValHisAsnLeuProGlnAsnLeu	
240	actggctacatctggtacaaaggggcaaagtacggacctctaccattacattacatcatat	299
	ThrGlyTyrIleTrpTyrLysGlyGlnMetThrAspLeuTyrHisTyrIleThrSerTyr	
300	gtagtagacgggtcaaattatataatgggcctgcctacagtggacgagaaacagtatatcc	359
40	ValValAspGlyGlnIleIleTyrGlyProAlaTyrSerGlyArgGluThrValTyrSer	
360	aatgcatccctgctgatccagaatgtcacacaggaggatgcaggatccctacaccttacac	419
	AsnAlaSerLeuLeuIleGlnAsnValThrGlnGluAspAlaGlySerTyrThrLeuHis	
420	atcataaagcgaggcgatgggactggaggagtaactggatatttcactgtcaccttatac	479
45	IleIleLysArgGlyAspGlyThrGlyGlyValThrGlyTyrPheThrValThrLeuTyr	
480	tcggagactcccaagcgtccatctccagcagcaacttaaaccgccaggagggtcatggag	539
	SerGluThrProLysArgSerIleSerSerSerAsnLeuAsnProArgGluValMetGlu	

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	540	gctgtgcgcttaaatctgtgatcctgagactccggatgcaagctacctgtggttgctgaat AlaValArgLeuIleCysAspProGluThrProAspAlaSerTyrLeuTrpLeuLeuAsn	599
5	600	ggtcagaacctccctatgactcacaggttgagctgtccaaaaccaacaggaccctctat GlyGlnAsnLeuProMetThrHisArgLeuGlnLeuSerLysThrAsnArgThrLeuTyr	659
	660	ctatttggtgtcacaaagtatatattgcagggcctatgaatgtgaaatacggaggggagtg LeuPheGlyValThrLysTyrIleAlaGlyProTyrGluCysGluIleArgArgGlyVal	719
10	720	agtgccagccgcagtgaccagtcaccctgaatctcctcccgaagctgccatgccttac SerAlaSerArgSerAspProValThrLeuAsnLeuLeuProLysLeuProMetProTyr	779
	780	atcaccatcaacaacttaaaccacagggagaagaaggatgtgttagccttcacctgtgaa IleThrIleAsnAsnLeuAsnProArgGluLysLysAspValLeuAlaPheThrCysGlu	839
15	840	cctaagagtcggaactacacctacatttggtggctaaatggtcagagcctcccggtcagt ProLysSerArgAsnTyrThrTyrIleTrpTrpLeuAsnGlyGlnSerLeuProValSer	899
	900	ccgagggtaaaagcgacccattgaaaacaggatactcattctaccagtgctcagagaaat ProArgValLysArgProIleGluAsnArgIleLeuIleLeuProSerValThrArgAsn	959
20	960	gaaacaggaccctatcaatgtgaaatacgggaccgatatggtggcatccgcagtaacca GluThrGlyProTyrGlnCysGluIleArgAspArgTyrGlyGlyIleArgSerAsnPro	1019
	1020	gtcacctgaatgtcctctatgggtccagacctcccagaatttacccttacttcacctat ValThrLeuAsnValLeuTyrGlyProAspLeuProArgIleTyrProTyrPheThrTyr	1079
25	1080	taccgttcaggagaaaaacctcgacttgctctgtttgaggactctaaccaccggcagag TyrArgSerGlyGluAsnLeuAspLeuSerCysPheAlaAspSerAsnProProAlaGlu	1139
	1140	tatTTTTGGACAATTAATGGGAAGTTTCAGCTATCAGGACAAAAGCTCTTTATCCCCAA TyrPheTrpThrIleAsnGlyLysPheGlnLeuSerGlyGlnLysLeuPheIleProGln	1199
30	1200	attactacaaatcatagcgggctctatgcttgctctgttcgtaactcagccactggcaag IleThrThrAsnHisSerGlyLeuTyrAlaCysSerValArgAsnSerAlaThrGlyLys	1259
	1260	gaaatctccaaatccatgatagtcaaagtctctgggtccctggcatggaaaccagacagag GluIleSerLysSerMetIleValLysValSerGlyProCysHisGlyAsnGlnThrGlu	1319
35	1320	tctcattaatggctgccacaatagagacactgagaaaaagaacagggttgataccttcag SerHisEnd	1379
	1380	aaattcaagacaaagaagaaaaaggctcaatgttattggactaaataatcaaaaggataa	1439
	1440	tgTTTTcataatTTTTattggaaaatgtgctgattcttggaatgttttattctccagatt	1499
	1500	tatgaactTTTTtcttcagcaattggtaaagtatactTTTgtaaacaaaaattgaaaca	1559
	1560	TTTgctTTTTgctctctatctgagtgccccccc 1591	

It will be appreciated that the instant specification and claims are set forth by way of illustration and not limitation and that various modifications and changes may be made without departing from the scope of the present invention.

Claims

1. A nucleic acid comprising a base sequence which codes for a peptide sequence, characterized in that the group nucleic acid is a DNA selected from the following group of five sequences:

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10 30 50
CAGCCGTGCTCGAAGCGTTCCTGGAGCCCAAGCTCTCCTCCACAGGTGAAGACAGGGCCA

5 70 90 110
GCAGGAGACACCATGGGGGCACCTCTCAGCCCCACTTCACAGAGTGCCTGTACCCTGGCAG
MetGlyHisLeuSerAlaProLeuHisArgValArgValProTrpGln

10 130 150 170
GGGCTTCTGCTCAGAGCCTCACTTCTAACCCTTCTGGAACCCGCCCACCACTGCCCAGCTC
15 GlyLeuLeuLeuThrAlaSerLeuLeuThrPheTrpAsnProProThrThrAlaGlnLeu

190 210 230
20 ACTACTGAATCCATGCCATTCAATGTTGCAGAGGGGAAGGAGGTCTTCTCCTTGTCAC
ThrThrGluSerMetProPheAsnValAlaGluGlyLysGluValLeuLeuLeuValHis

250 270 290
AATCTGCCCCAGCAACTTTTGGCTACAGCTGGTACAAAGGGGAAAGAGTGGATGGCAAC
25 AsnLeuProGlnGlnLeuPheGlyTyrSerTrpTyrLysGlyGluArgValAspGlyAsn

310 330 350
30 CGTCAAATTGTAGGATATGCAATAGGAACTCAACAAGCTACCCCAGGGCCCGCAAACAGC
ArgGlnIleValGlyTyrAlaIleGlyThrGlnGlnAlaThrProGlyProAlaAsnSer

370 390 410
35 GGTCGAGAGACAATATACCCCAATGCATCCCTGCTGATCCAGAACGTCACCCAGAATGAC
GlyArgGluThrIleTyrProAsnAlaSerLeuLeuIleGlnAsnValThrGlnAsnAsp

430 450 470
40 ACAGGATTCTACACCCTACAAGTCATAAAGTCAGATCTTGTGAATGAAGAAGCAACTGGA
ThrGlyPheTyrThrLeuGlnValIleLysSerAspLeuValAsnGluGluAlaThrGly

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490 510 530
 CAGTTCCATGTATACCCGGAGCTGCCCAAGCCCTCCATCTCCAGCAACAACCTCCAACCCCT
 5 GlnPheHisValTyrProGluLeuProLysProSerIleSerSerAsnAsnSerAsnPro

550 570 590
 10 GTGGAGGACAAGGATGCTGTGGCCTTCACCTGTGAACCTGAGACTCAGGACACAACCTAC
 ValGluAspLysAspAlaValAlaPheThrCysGluProGluThrGlnAspThrThrTyr

610 630 650
 15 CTGTGGTGGATAAACAATCAGAGCCTCCCGGTGAGTCCCAGGCTGCAGCTGTCCAATGGC
 LeuTrpTrpIleAsnAsnGlnSerLeuProValSerProArgLeuGlnLeuSerAsnGly

670 690 710
 20 AACAGGACCCTCACTCTACTCAGTGTCAACAAGGAATGACACAGGACCCTATGAGTGTGAA
 AsnArgThrLeuThrLeuLeuSerValThrArgAsnAspThrGlyProTyrGluCysGlu

730 750 770
 25 ATACAGAACCCAGTGAGTGCGAACCAGTACCCAGTCACCTTGAATGTCACCTATGGC
 IleGlnAsnProValSerAlaAsnArgSerAspProValThrLeuAsnValThrTyrGly

790 810 830
 30 CCGGACACCCCCACCATTTCCCTTCAGACACCTATTACCGTCCAGGGGGCAACCTCAGC
 ProAspThrProThrIleSerProSerAspThrTyrTyrArgProGlyAlaAsnLeuSer

850 870 890
 35 CTCTCCTGCTATGCAGCCTCTAACCACCTGCACAGTACTCCTGGCTTATCAATGGAACA
 LeuSerCysTyrAlaAlaSerAsnProProAlaGlnTyrSerTrpLeuIleAsnGlyThr

910 930 950
 45 TTCCAGCAAAGCACACAAGAGCTCTTTATCCCTAACATCACTGTGAATAATAGTGGATCC
 PheGlnGlnSerThrGlnGluLeuPheIleProAsnIleThrValAsnAsnSerGlySer

970 990 1010
 50 TATACCTGCCACGCCAATAACTCAGTCACTGGCTGCAACAGGACCACAGTCAAGACCGATC
 TyrThrCysHisAlaAsnAsnSerValThrGlyCysAsnArgThrThrValLysThrIle

55

1030 1050 1070
5 ATAGTCACTGATAATGCTCTACCACAAGAAAATGGCCTCTCACCTGGGGCCATTGCTGGC
IleValThrAspAsnAlaLeuProGlnGluAsnGlyLeuSerProGlyAlaIleAlaGly

1090 1110 1130
10 ATTGTGATTGGAGTAGTGGCCCTGGTTGCTCTGATAGCAGTAGCCCTGGCATGTTTTCTG
IleValIleGlyValValAlaLeuValAlaLeuIleAlaValAlaLeuAlaCysPheLeu

1150 1170 1190
15 CATTTCCGGGAAGACCGGCAGGGCAAGCGACCAGCGTGATCTCACAGAGCACAAACCCTCA
HisPheGlyLysThrGlyArgAlaSerAspGlnArgAspLeuThrGluHisLysProSer

1210 1230 1250
20 GTCTCCAAGCACACTCAGGACCACTCCAATGACCCACCTAACAAGATGAATGAAGTTACT
ValSerAsnHisThrGlnAspHisSerAsnAspProProAsnLysMetAsnGluValThr

1270 1290 1310
25 TATTCTACCTGAACTTTGAAGCCAGCAACCCACACAACCAACTTCAGCCTCCCCATCC
TyrSerThrLeuAsnPheGluAlaGlnGlnProThrGlnProThrSerAlaSerProSer

1330 1350 1370
30 CTAACAGCCACAGAAATAATTTATTCAGAAAGTAAAAAGCAGTAATGAAACCTGTCCTGC
LeuThrAlaThrGluIleIleTyrSerGluValLysLysGln

1390 1410 1430
35 TCACTGCAGTGCTGATGTATTTCAAGTCTCTCACCTCATCACTAGGAGATTCTTTCCC

1450 1470 1490
40 CTGTAGGGTAGAGGGGTGGGGACAGAAACAACCTTTCTCCTACTCTTCCTTCCTAATAGGC

1510 1530 1550
45 ATCTCCAGGCTGCCTGGTCACTGCCCCCTCTCTCAGTGTCAAATAGATGAAAGTACATTGGG

1570 1590 1610
50 AGTCTGTAGGAAACCCAACCTTCTTGTCATTGAAATTGGCAAAGCTGACTTTGGGAAAG

1630 1650 1670
AGGGACCAGAACTTCCCCTCCCTTCCCCTTTTCCCAACCTGGACTTGTTTTAAACTTGCC
5

1690 1710 1730
TGTTTCAGAGCACTCATTCCCTTCCCACCCCCAGTCCTGTCCTATCACTCTAATTTCGGATTT
10

1750 1770 1790
GCCATAGCCTTGAGGTTATGTCCTTTTCCATTAAGTACATGTGCCAGGAAACAGCGAGAG
15

1810 1830 1850
AGAGAAAGTAAACGGCAGTAATGCTTCTCCTATTTCTCCAAAGCCTTGTGTGAAGTAGCA
20

1870 1890 1910
AAGAGAAGAAAATCAAATATATAACCAATAGTGAAATGCCACAGGTTTGTCCACTGTCAG
25

1930 1950 1970
GGTTGTCTACCTGTAGGATCAGGGTCTAAGCACCTTGGTGCTTAGCTAGAAATACCACCTA
30

1990 2010 2030
ATCCTTCTGGCAAGCCTGTCTTCAGAGAACCCACTAGAAAGCAACTAGGAAAAATCACTTG
35

2050 2070 2090
CCAAAATCCAAGGCAATTCTTGATGGAAAAATGCAAAAGCACATATATGTTTTAATATCTT
40

2110 2130 2150
TATGGGCTCTGTTCAGGCAAGTGCCTGAGAGGGAGGGGTTATAGCTTCAGGAGGGAACCAAG
45

2170 2190 2210
CTTCTGATAAAACACAATCTGCTAGGAACTTGGGAAAGGAATCAGAGAGCTGCCCTTCAGC
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2230 2250 2270
 GATTATTTAAATTGTTAAAGAATACACAATTTGGGGTATTGGGATTTTTCTCCTTTTCTC
 5
 2290 2310 2330
 TGAGACATTGCAECATTTTAAATTTTGTAACTGCTTATTTATGTGAAAAGGGTTATTTT...
 10
 2350 2370 2390
 ACTTAGCTTAGCTATGTCAGCCAATCCGATTGCCTTAGGTGAAAGAAACCACCGAAATCC
 15
 2410 2430 2450
 CTCAGGTCCCTTGGTCAGGAGCCTCTCAAGATTTTTTTTGTGAGAGGCTCCAAATAGAAA
 20
 2470 2490 2510
 ATAAGAAAAGGTTTTCTTCATTCATGGCTAGAGCTAGATTTAACTCAGTTTCTAGGCACC
 25
 2530 2550 2570
 TCAGACCAATCATCAACTACCATTTCTATTCATGTTTGCACCTGTGCATTTTCTGTTTGC
 30
 2590 2610 2630
 CCCCATTCACTTTGTGAGGAAACCTTGGCCTCTGCTAAGGTGTATTTGGTCCTTGAGAAG
 35
 2650 2670 2690
 TGGGAGCACCTTACAGGGACACTATCACTCATGCTGGTGGCATTGTTTACAGCTAGAAAG
 40
 2710 2730 2750
 CTGCAC TGGTGTCTAATGCCCTTGGGAAATGGGGCTGTGAGGAGGAGGATTATAACTTAG
 45
 2770 2790 2810
 GCCTAGCCTCTTTTAAACAGCCTCTGAAATTTATCTTTTCTTCTATGGGGTCTATAAATGT
 50
 2830 2850 2870
 ATCTTATAAATAAAAAGGAAGGACAGGAGGAAGACAGGCAATGTACTTCTCACCACGTCT
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2890 2910 2930
TCTACACAGATGGAATCTCTTTGGGGCTAAGAGAAAGGTTTTATTCTATATTGCTTACCT
5
2950 2970 2990
GATCTCATGTTAGGCCTAAGAGGCTTTCCTCCAGGAGGATTAGCTTGGAGTTCTCTATACT
10
3010 3030 3050
CAGGTACCTCTTTCAGGGTTTTCTAACCCTGACACGGACTGTGCATACTTTCCTCATCC
15
3070 3090 3110
ATGCTGTGCTGTGTTATTTAATTTTCTGGCTAAGATCATGTCTGAATTATGTATGAAA
20
3130 3150 3170
ATTATTCTATGTTTTATAATAAAAATAATATATCAGACATCGAAAAAAAAAA,
25
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35
40
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(2)

5 10 30 50
 CAGCCGTGCTCGAAGCGTTCTCTGGAGCCCAAGCTCTCCTCCACAGGTGAAGACAGGGCCA

 10 70 90 110
 GCAGGAGACACCATGGGGCACCTCTCAGCCCCACTTCACAGAGTGCGTGTTACCCTGGCAG
 MetGlyHisLeuSerAlaProLeuHisArgValArgValProTrpGln

 15 130 150 170
 GGGCTTCTGCTCACAGCCTCACTTCTAACCCTTCTGGAACCCGCCCACCACTGCCCAGCTC
 GlyLeuLeuLeuThrAlaSerLeuLeuThrPheTrpAsnProProThrThrAlaGlnLeu

 20 190 210 230
 ACTACTGAATCCATGCCATTCAATGTTGCAGAGGGGAAGGAGGTCTTCTCCTTGTCCAC
 ThrThrGluSerMetProPheAsnValAlaGluGlyLysGluValLeuLeuLeuValHis

 25 250 270 290
 AATCTGCCCCAGCAACTTTTTGGCTACAGCTGGTACAAAGGGGAAAGAGTGGATGGCAAC
 30 AsnLeuProGlnGlnLeuPheGlyTyrSerTrpTyrLysGlyGluArgValAspGlyAsn

 310 330 350
 35 CGTCAAATTGTAGGATATGCAATAGGAACTCAACAAGCTACCCCAGGGCCCGCAAACAGC
 ArgGlnIleValGlyTyrAlaIleGlyThrGlnGlnAlaThrProGlyProAlaAsnSer

 370 390 410
 40 GGTCGAGAGACAATATACCCCAATGCATCCCTGCTGATCCAGAACGTCACCCAGAATGAC
 GlyArgGluThrIleTyrProAsnAlaSerLeuLeuIleGlnAsnValThrGlnAsnAsp

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EP 0 346 710 B1

430 450 470
ACAGGATTCTACACCTACAAGTCATAAAGTCAGATCTTGTGAATGAAGAAGCAACTGGA
5 ThrGlyPheTyrThrLeuGlnValIleLysSerAspLeuValAsnGluGluAlaThrGly

490 510 530
10 CAGTTCCATGTATACCCGGAGCTGCCCAAGCCCTCCATCTCCAGCAACAACCTCCAACCT
GlnPheHisValTyrProGluLeuProLysProSerIleSerSerAsnAsnSerAsnPro

550 570 590
15 GTGGAGGACAAGGATGCTGTGGCCTTCACCTGTGAACCTGAGACTCAGGACACAACCTAC
ValGluAspLysAspAlaValAlaPheThrCysGluProGluThrGlnAspThrThrTyr

610 630 650
20 CTGTGGTGGATAAACAATCAGAGCCTCCCGGTCAGTCCCAGGCTGCAGCTGTCCAATGGC
LeuTrpTrpIleAsnAsnGlnSerLeuProValSerProArgLeuGlnLeuSerAsnGly

670 690 710
25 AACAGGACCCCTCACTCTACTCAGTGTACAAAGGAATGACACAGGACCCTATGAGTGTGAA
AsnArgThrLeuThrLeuLeuSerValThrArgAsnAspThrGlyProTyrGluCysGlu

730 750 770
30 ATACAGAACCAGTGAGTGCGAACCGCAGTGACCCAGTCACCTTGAATGTCACCTATGGC
IleGlnAsnProValSerAlaAsnArgSerAspProValThrLeuAsnValThrTyrGly

790 810 830
35 CCGGACACCCCAACCATTTCCCCTTCAGACACCTATTACCGTCCAGGGGCAAACCTCAGC
40 ProAspThrProThrIleSerProSerAspThrTyrTyrArgProGlyAlaAsnLeuSer

45

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850 870 890
 CTCTCTGCTATGCAGCCTCTAACCACCTGCACAGTACTCCTGGCTTATCAATGGAACA
 5 LeuSerCysTyrAlaAlaSerAsnProProAlaGlnTyrSerTrpLeuIleAsnGlyThr

910 930 950
 10 TTCCAGCAAAGCACACAAGAGCTCTTTATCCCTAACATCACTGTGAATAATAGTGGATCC
 PheGlnGlnSerThrGlnGluLeuPheIleProAsnIleThrValAsnAsnSerGlySer

970 990 1010
 15 TATACCTGCCACGCCAATAACTCAGTCACTGGCTGCAACAGGACCACAGTCAAGACGATC
 TyrThrCysHisAlaAsnAsnSerValThrGlyCysAsnArgThrThrValLysThrIle

1030 1050 1070
 20 ATAGTCACTGAGCTAAGTCCAGTAGTAGCAAAGCCCCAAATCAAAGCCAGCAAGACCACA
 IleValThrGluLeuSerProValValAlaLysProGlnIleLysAlaSerLysThrThr

1090 1110 1130
 25 GTCACAGGAGATAAGGACTCTGTGAACCTGACCTGCTCCACAAATGACACTGGAATCTCC
 ValThrGlyAspLysAspSerValAsnLeuThrCysSerThrAsnAspThrGlyIleSer

1150 1170 1190
 30 ATCCGTTGGTTCTTCAAAAACCAGAGTCTCCCGTCCTCGGAGAGGATGAAGCTGTCCAG
 IleArgTrpPhePheLysAsnGlnSerLeuProSerSerGluArgMetLysLeuSerGln

1210 1230 1250
 35 GGCAACACCACCCTCAGCATAAACCCTGTCAAGAGGGAGGATGCTGGGACGTATTGGTGT
 40 GlyAsnThrThrLeuSerIleAsnProValLysArgGluAspAlaGlyThrTyrTrpCys

45

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1270 1290 1310
5 GAGGTCTTCAACCCAATCAGTAAGAACCAAAGCGACCCCATCATGCTGAACGTAAACTAT
GluValPheAsnProIleSerLysAsnGlnSerAspProIleMetLeuAsnValAsnTyr

1330 1350 1370
10 AATGCTCTACCAACAAGAAAATGGCCTCTCACCTGGGGCCATTGCTGGCATTGTGATTGGA
AsnAlaLeuProGlnGluAsnGlyLeuSerProGlyAlaIleAlaGlyIleValIleGly

1390 1410 1430
15 GTAGTGGCCCTGGTTGCTCTGATAGCAGTAGCCCTGGCATGTTTTCTGCATTTCTGGGAAG
ValValAlaLeuValAlaLeuIleAlaValAlaLeuAlaCysPheLeuHisPheGlyLys

1450 1470 1490
20 ACCGGCAGCTCAGGACCACTCCAATGACCCACCTAACAAGATGAATGAAGTTACTTATTC
ThrGlySerSerGlyProLeuGln

1510 1530 1550
25 TACCCTGAACTTTGAAGCCCAGCAACCCACACAACCAACTTCAGCCTCCCCATCCCTAAC

1570 1590 1610
30 AGCCACAGAAATAATTTATTCAGAAGTAAAAAAGCAGTAATGAAACCTGAAAAA

1630
35 AAAAAAAAAA

40

45

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(3)

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CAGCCGTGCTCGAAGCGTTCCTGGAGCCCAAGCTCTCCTCCACAGGTGAACACAGGGCCA

10

70

90

110

GCAGGAGACACCATGGGGCACCTCTCAGCCCCACTTCACAGAGTGCGTGTACCCTGGCAG
MetGlyHisLeuSerAlaProLeuHisArgValArgValProTrpGln

15

130

150

170

GGGCTTCTGCTCACAGCCTCACTTCTAACCTTCTGGAACCCGCCCACCACTGCCCAGCTC
GlyLeuLeuLeuThrAlaSerLeuLeuThrPheTrpAsnProProThrThrAlaGlnLeu

20

190

210

230

ACTACTGAATCCATGCCATTCAATGTTGCAGAGGGGAAGGAGGTTCTTCTCCTTGTCCAC
ThrThrGluSerMetProPheAsnValAlaGluGlyLysGluValLeuLeuLeuValHis

25

250

270

290

AATCTGCCCCAGCAACTTTTGGCTACAGCTGGTACAAAGGGGAAAGAGTGGATGGCAAC
AsnLeuProGlnGlnLeuPheGlyTyrSerTrpTyrLysGlyGluArgValAspGlyAsn

30

310

330

350

CGTCAAATTGTAGGATATGCAATAGGAACTCAACAAGCTACCCCAGGGCCCGCAAACAGC
ArgGlnIleValGlyTyrAlaIleGlyThrGlnGlnAlaThrProGlyProAlaAsnSer

35

370

390

410

GGTCGAGAGACAATATACCCCAATGCATCCCTGCTGATCCAGAACGTCACCCAGAAATGAC
GlyArgGluThrIleTyrProAsnAlaSerLeuLeuIleGlnAsnValThrGlnAsnAsp

40

430

450

470

ACAGGATTCTACACCCTACAAGTCATAAAGTCAGATCTTGTGAATGAAGAAGCAACTGGA
ThrGlyPheTyrThrLeuGlnValIleLysSerAspLeuValAsnGluGluAlaThrGly

50

55

490 510 530
 CAGTTCATGTATACCCGGAGCTGCCCAAGCCCTCCATCTCCAGCAACAACCTCCAACCT
 5 GlnPheHisValTyrProGluLeuProLysProSerIleSerSerAsnAsnSerAsnPro

550 570 590
 10 GTGGAGGACAAGGATGCTGTGGCCTTCACCTGTGAACCTGAGACTCAGGACACAACCTAC
 ValGluAspLysAspAlaValAlaPheThrCysGluProGluThrGlnAspThrThrTyr

610 630 650
 15 CTGTGGTGGATAAACAATCAGAGCCTCCCGGTCAGTCCCAGGCTGCAGCTGTCCAATGGC
 LeuTrpTrpIleAsnAsnGlnSerLeuProValSerProArgLeuGlnLeuSerAsnGly

670 690 710
 20 AACAGGACCCTCACTCTACTCAGTGTCAACGAATGACACAGGACCCTATGAGTGTGAA
 AsnArgThrLeuThrLeuLeuSerValThrArgAsnAspThrGlyProTyrGluCysGlu

730 750 770
 25 ATACAGAACCCAGTGAGTGCGAACCGCAGTGACCCAGTCACCTTGAATGTCACCTATGGC
 IleGlnAsnProValSerAlaAsnArgSerAspProValThrLeuAsnValThrTyrGly

790 810 830
 30 CCGGACACCCCCACCATTTCCECTTCAGACACCTATTACCGTCCAGGGGCAAACCTCAGC
 35 ProAspThrProThrIleSerProSerAspThrTyrTyrArgProGlyAlaAsnLeuSer

850 870 890
 40 CTCTCCTGCTATGCAGCCTCTAACCCACCTGCACAGTACTCCTGGCTTATCAATGGAACA
 LeuSerCysTyrAlaAlaSerAsnProProAlaGlnTyrSerTrpLeuIleAsnGlyThr

910 930 950
 45 TTCCAGCAAAGCACACAAGAGCTCTTTATCCCTAACATCACTGTGAATAATAGTGGATCC
 PheGlnGlnSerThrGlnGluLeuPheIleProAsnIleThrValAsnAsnSerGlySer

970 990 1010
 50 TATACCTGCCACGCCAATAACTCAGTCACTGGCTGCAACAGGACCACAGTCAAGACGATC
 TyrThrCysHisAlaAsnAsnSerValThrGlyCysAsnArgThrThrValLysThrIle

55

	1030	1050	1070
5	ATAGTCACTGATAATGCTCTACCACAAGAAAATGGCCTCTCACCTGGGGCCATTGCTGGC IleValThrAspAsnAlaLeuProGlnGluAsnGlyLeuSerProGlyAlaIleAlaGly		
	1090	1110	1130
10	ATTGTGATTGGAGTAGTGGCCCTGGTTCCTCTGATAGCAGTAGCCCTGGCATGTTTTCTG IleValIleGlyValValAlaLeuValAlaLeuIleAlaValAlaLeuAlaCysPheLeu		
	1150	1170	1190
15	CATTTCCGGGAAGACCGGCAGCTCAGGACCACTCCAATGACCCACCTAACAAAGATGAATGA HisPheGlyLysThrGlySerSerGlyProLeuGln		
	1210	1230	1250
20	AGTTACTTATTCTACCCTGAACTTTGAAGCCCAGCAACCCACACAACCAACTTCAGCCTC		
	1270	1290	1310
25	CCCATCCCTAACAGCCACAGAAATAATTTATTTCAGAAGTAAAAAAGCAGTAATGAAACCT		
	1330		
30	GAAAAAAAAAAAAAAAAAAAA		

35

40

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(4)

5	1	acagcacagctgacagccgtactcaggaagcttctggatcctaggcttatctccacagag	60
	61	gagaacacacaagcagcagagaccatggggccctctcagccctccctgcacacacctc MetGlyProLeuSerAlaProProCysThrHisLeu	120
10	121	atcacttgggaaggggtcctgctcacagcatcacttttaacttctggaatccgcccaca... IleThrTrpLysGlyValLeuLeuThrAlaSerLeuLeuAsnPheTrpAsnProProThr	180
	181	actgcccgaagtcacgattgaagcccagccacccaaagtcttgaggggaaggatgttctt ThrAlaGlnValThrIleGluAlaGlnProProLysValSerGluGlyLysAspValLeu	240
15	241	ctacttgtccacaatttgccccagaatcttgctggctacatttggtacaaagggcaaagt LeuLeuValHisAsnLeuProGlnAsnLeuAlaGlyTyrIleTrpTyrLysGlyGlnMet	300
	301	acatacgtctaccattacattacatcatatgtagtagacggtcaaagaattatataatggg ThrTyrValTyrHisTyrIleThrSerTyrValValAspGlyGlnArgIleIleTyrGly	360
20	361	cctgcatacagtggaagagaaagagtataattccaatgcatccctgctgätcagaaatgtc ProAlaTyrSerGlyArgGluArgValTyrSerAsnAlaSerLeuLeuIleGlnAsnVal	420
	421	acgcaggaggatgcaggatcctacaccttacacatcataaagcgäcgcgätgggäactgga ThrGlnGluAspAlaGlySerTyrThrLeuHisIleIleLysArgArgAspGlyThrGly	480
	481	ggagtaactggacatttcaccttcaccttacacctggagactcccaagccctccatctcc GlyValThrGlyHisPheThrPheThrLeuHisLeuGluThrProLysProSerIleSer	540
25	541	agcagcaacttaaatcccagggaggccatggaggctgtgatcttaacctgtgatcctgcg SerSerAsnLeuAsnProArgGluAlaMetGluAlaValIleLeuThrCysAspProAla	600
	601	actccagccgcaagctaccagtgggtggatgaatggctcagagcctccctatgactcacagg ThrProAlaAlaSerTyrGlnTrpTrpMetAsnGlyGlnSerLeuProMetThrHisArg	660
30	661	ttgcagctgtccaaaaccaacaggaccctctttatatttggtgtcacaagtatattgca LeuGlnLeuSerLysThrAsnArgThrLeuPheIlePheGlyValThrLysTyrIleAla	720
	721	ggaccctatgaatgtgaaatacggaaaccagtgagtgccagccgäcagtgacccagtcacc GlyProTyrGluCysGluIleArgAsnProValSerAlaSerArgSerAspProValThr	780
35	781	ctgaatctcctcccaaagctgtccaagccctacatcacaaatcaacaacttaaacccaga LeuAsnLeuLeuProLysLeuSerLysProTyrIleThrIleAsnAsnLeuAsnProArg	840
	841	gagaataaggatgtcttaaccttcacctgtgaacctaaagtgagaactacacctacatt GluAsnLysAspValLeuThrPheThrCysGluProLysSerGluAsnTyrThrTyrIle	900
40	901	tgggtggctaaatggctcagagcctccctgtcagtcacccagggtaaagcgacccattgaaaac TrpTrpLeuAsnGlyGlnSerLeuProValSerProArgValLysArgProIleGluAsn	960
	961	aggatcctcattctacccaatgtcacgagaaatgaaacaggaccttatcaatgtgaaata ArgIleLeuIleLeuProAsnValThrArgAsnGluThrGlyProTyrGlnCysGluIle	1020
45	1021	cgggaccgatattggtggcatccgäcagtgacccagtcacccctgaatgtcctctatggtcca ArgAspArgTyrGlyGlyIleArgSerAspProValThrLeuAsnValLeuTyrGlyPro	1080

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1 ggggtggatcctagggtcatctccataggggagaacacacatacagcagagaccatggga 59
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 20

2. A replicable recombinant cloning vehicle having an insert comprising a nucleic acid of claim 1.

25 3. A cell that is transfected, infected or injected with a recombinant cloning vehicle of claim 2.

4. A method for preparing a polypeptide, said method comprising the steps of

(a) culturing the cell of claim 3

(b) recovering the polypeptide expressed by said cell.

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5. A method for preparing an antibody directed against a polypeptide said method comprising the steps of

(a) preparing said polypeptide by the method of claim 4

(b) injecting said polypeptide into a host capable of producing antibodies and

(c) recovering said antibodies.

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Patentansprüche

1. Nucleinsäure, umfassend eine Basen-Sequenz, die für eine Peptid-Sequenz codiert,
 dadurch gekennzeichnet, daß die Gruppen-Nucleinsäure eine DNA ist, die aus der folgenden Gruppe
 40 von fünf Sequenzen ausgewählt ist:

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490 510 530
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 5 GlnPheHisValTyrProGluLeuProLysProSerileSerSerAsnAsnSerAsnPro

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 10 ValGluAspLysAspAlaValAlaPheThrCysGluProGluThrGlnAspThrThrTyr

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(4)

5 1 acagcacagctgacagccgtactcaggaagcttctggatcctaggcttatctccacagag 60
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 121 atcacttggaaaggggtcctgctcagcagcatcacttttaaacttctggaaatccgcccaca. 180
 10 IleThrTrpLysGlyValLeuLeuThrAlaSerLeuLeuAsnPheTrpAsnProProThr
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 ThrAlaGlnValThrIleGluAlaGlnProProLysValSerGluGlyLysAspValLeu
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 15 LeuLeuValHisAsnLeuProGlnAsnLeuAlaGlyTyrIleTrpTyrLysGlyGlnMet
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 20 ProAlaTyrSerGlyArgGluArgValTyrSerAsnAlaSerLeuLeuIleGlnAsnVal
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 25 541 agcagcaacttzaatcccagggaggccatggagsgtgtgatcttaacctgtgatcctgcg 600
 SerSerAsnLeuAsnProArgGluAlaMetGluAlaValIleLeuThrCysAspProAla
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 30 561 ttgcagctgtccaaaaccaacaggaccctcttttatatttggtgtcacaagtatattgca 720
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 841 gagaataaggatgtcttaaccttcacctgtgaacctagagtgagaactacacctacatt 900
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 45 1021 cgggaccgatatgggtggcatccgcagtgaccagtcaccctgaatgtcctctatggtcca 1080
 ArgAspArgTyrGlyGlyIleArgSerAspProValThrLeuAsnValLeuTyrGlyPro

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1141	tcctgcttcggtgagtcctaaccacgggcacaaatattcttggacaattaatgggaagttt	1200
5	SerCysPheGlyGluSerAsnProArgAlaGlnTyrSerTrpThrIleAsnGlyLysPhe	
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	GlnLeuSerGlyGlnLysLeuSerIleProGlnIleThrThrLysHisSerGlyLeuTyr	
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10	AlaCysSerValArgAsnSerAlaThrGlyLysGluSerSerLysSerIleThrValLys	
1321	gtctctgactggatattaccctgaattctactagtctcctccaattccattttctcccatg	1380
	ValSerAspTrpIleLeuProEnd	
1381	gaatcacgaagagcaagacccactctgttccagaagccctataatctggaggtggacaac	1440
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1441	aaaatgttcgacaccataacaacagctactcaaactgtaaaccaggataagaagttgatg	1560
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1561	ctccaccctctactgtctgtctcatgcctgcctctttcacttggcaggataatgcagtcac	1680
1621	tagaatttcacatgtagtagcttctgagggtacaacagagtgtcagatatgtcatctca	1740
1681	acctcaaacttttcgtaacatctcagggaaatgtggctctctccatcttgcatacaggg	1800
1741	ctcccaatagaaatgaaacacagagatattgcctgtgtgtttgcagagaagatgggttcta	1860
20	tazagagtaggaaagctgaaattatagtagagtctcctttaaatgcacattgtgtggatg	1920
1801	gctctcaccatttcctaagagatacagtgtaaaacggtgacagtaatactgattctagca	1980
1861	gaaataaacatgtaccacatttgcaaaaaa	2010
1921		
1981		

end

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(5)

1 ggggtggatcctagggtcatctccatagggggagaacacacatacagcagagaccatggga 59
 MetGly
 50 cccctctcagccctccctgcactcagcacatcacctggaaggggctcctgctcacagca 119
 ProLeuSerAlaProProCysThrGlnHisIleThrTrpLysGlyLeuLeuLeuThrAla
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 SerLeuLeuAsnPheTrpAsnLeuProThrThrAlaGlnValIleIleGluAlaGlnPro
 180 cccaaagtcttctgaggggaaggatgttcttctacttgtccacaatttgccccagaatctt 239
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 360 aatgcatccctgctgatccagaatgtcacacaggaggatgcaggatcctacaccttacac 419
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 420 atcataaagcgaggcgatgggactggaggagtaactggatatttcactgtcaccttatac 479
 IleIleLysArgGlyAspGlyThrGlyGlyValThrGlyTyrPheThrValThrLeuTyr
 480 tcggagactcccaagcgctccatctccagcagcaacttaaacccccaggaggtcatggag 539
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 AlaValArgLeuIleCysAspProGluThrProAspAlaSerTyrLeuTrpLeuLeuAsn
 600 ggtcagaacctccctatgactcacaggttgagctgtccaaaaccaacaggaccctctat 659
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 660 ctatattggtgtcacaaagtatatgtcagggccctatgaatgtgaaatacggaggggagtg 719
 LeuPheGlyValThrLysTyrIleAlaGlyProTyrGluCysGluIleArgArgGlyVal
 720 agtgccagccgcagtgacctcagtcacctgaatctcctcccgaagctgcccattgccttac 779
 SerAlaSerArgSerAspProValThrLeuAsnLeuLeuProLysLeuProMetProTyr
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 IleThrIleAsnAsnLeuAsnProArgGluLysLysAspValLeuAlaPheThrCysGlu
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 ProArgValLysArgProIleGluAsnArgIleLeuIleLeuProSerValThrArgAsn
 960 gaaacaggaccctatcaatgtgaaatacgggaccgatatgggtggcatccgcagtaaccca 1019
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1020 gtcaccctgaatgtcctctatgggtccagacctccccagaattttacccttacttcacctat 1079
 ValThrLeuAsnValLeuTyrGlyProAspLeuProArgIleTyrProTyrPheThrTyr
 1080 taccgttcaggagaaaaacctcgacttgtcctgctttgaggactctaaccaccgagag 1139
 5 TyrArgSerGlyGluAsnLeuAspLeuSerCysPheAlaAspSerAsnProProAlaGlu
 1140 tatTTTTggacaattaatgggaagtttcagctatcaggacaaaagctctttatcccccaa 1199
 TyrPheTrpThrIleAsnGlyLysPheGlnLeuSerGlyGlnLysLeuPheIleProGln
 1200 attactacaaatcatagcgggctctatgcttgctctgttcgtaactcagccactggcaag 1259
 10 IleThrThrAsnHisSerGlyLeuTyrAlaCysSerValArgAsnSerAlaThrGlyLys
 1260 gaaatctccaaatccatgatagtcaaaagtctctggtccctgccatggaaaccagacagag 1319
 GluIleSerLysSerMetIleValLysValSerGlyProCysHisGlyAsnGlnThrGlu
 1320 tctcattaatggctgccacaatagagacactgaagaaaaagaacagggttgataccttcatg 1379
 15 SerHisEnd
 1380 aaattcaagacazagaagaaaaaggctcaatgttattggactaaataatcaaaaggataa 1439
 1440 tgTTTTcataatttttattggaaaatgtgctgattcttggaatgttttattctccagatt 1499
 1500 tatgaactTTTTTTCTTCAGCAATTGGTAAAGTATACTTTGTAAACAAAAATTGAAACA 1559
 1560 tttgcttttgctctctatctgagtgccccccc 1591

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2. Replizierbares rekombinantes Kloniervehikel mit einem eine Nucleinsäure nach Anspruch 1 umfassenden Insert.
- 25 3. Zelle, die mit einem rekombinanten Kloniervehikel nach Anspruch 2 transfiziert, infiziert oder injiziert ist.
4. Verfahren zur Herstellung eines Polypeptids, umfassend die Schritte
 - (a) des Kultivierens der Zelle nach Anspruch 3,
 - (b) des Gewinnens des durch diese Zelle exprimierten Polypeptids.
- 30 5. Verfahren zur Herstellung eines gegen ein Polypeptid gerichteten Antikörpers, umfassend die Schritte
 - (a) des Herstellens des Polypeptids durch das Verfahren des Anspruchs 4,
 - (b) des Injizierens des Polypeptids in einen Wirt, der zur Bildung von Antikörpern befähigt ist, und
 - (c) des Gewinnens der Antikörper.

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Revendications

1. Acide nucléique comprenant une séquence de bases qui code pour une séquence peptidique, caractérisé en ce que le groupe d'acides nucléiques est de l'ADN choisi parmi le groupe de cinq séquences ci-après :

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10 30 50
 CAGCCGTCCTCGAAGCGTTCTCTGGAGCCCAAGCTCTCTCTCCACAGGTGAAGACAGGGCCA
 5
 70 90 110
 GCAGGAGACACCATGGGGCACCTCTCAGCCCCACTTCACAGAGTGCGTGTACCCTGGCAG
 10 MetGlyHisLeuSerAlaProLeuHisArgValArgValProTrpGln
 130 150 170
 GGGCTTCTGCTCAGACCTCACTTCTAACCCTTCTGGAACCCGCCCCACCACTGCCCCAGCTC
 15 GlyLeuLeuLeuThrAlaSerLeuLeuThrPheTrpAsnProProThrThrAlaGlnLeu
 190 210 230
 ACTACTGAATCCATGCCATTCGAATGTTGCAGAGGGGAAGGAGGTTCTTCTCCTTGCTCCAC
 20 ThrThrGluSerMetProPheAsnValAlaGluGlyLysGluValLeuLeuValHis
 250 270 290
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 25 AsnLeuProGlnGlnLeuPheGlyTyrSerTrpTyrLysGlyGluArgValAspGlyAsn
 310 330 350
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 30 ArgGlnIleValGlyTyrAlaIleGlyThrGlnGlnAlaThrProGlyProAlaAsnSer
 370 390 410
 GGTCCAGAGACAATATACCCCAATGCATCCCTGCTGATCCAGAACGTCACCCAGAAATGAC
 35 GlyArgGluThrIleTyrProAsnAlaSerLeuLeuIleGlnAsnValThrGlnAsnAsp
 430 450 470
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 40 ThrGlyPheTyrThrLeuGlnValIleLysSerAspLeuValAsnGluGluAlaThrGly
 45
 50
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490 510 530
 5 CAGTTCCATGTTATACCCGGAGCTGCCCAAGCCCTCCATCTCCAGCAACAACCTCCAACCCT
 GlnPheHisValTyrProGluLeuProLysProSerIleSerSerAsnAsnSerAsnPro

550 570 590
 10 GTGGAGGACAAAGCATGCTGTGGCCCTTCACCTGTGAACCTGAGACTCAGGACACAACTTAC
 ValGluAspLysAspAlaValAlaPheThrCysGluProGluThrGlnAspThrThrTyr

610 630 650
 15 CTGTGGTGGATAAACAATCAGAGCCCTCCCGGTCAGTCCCAGGCTGCAGCTGTCCAATGGC
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670 690 710
 20 AACAGGACCCTCACTCTACTCAGTGTCAACAAGGAATGACACAGGACCCTATGAGTGTCAA
 AsnArgThrLeuThrLeuLeuSerValThrArgAsnAspThrGlyProTyrGluCysGlu

730 750 770
 25 ATACAGAACCCAGTGAGTGGGAACCCGAGTGACCCAGTCACCTTGAATGTCACCTATGGC
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790 810 830
 30 CCGGACACCCCCACCATTTCCTCCCTTCAGACACCTATTACCGTCCAGGGGCAACCTCAGC
 ProAspThrProThrIleSerProSerAspThrTyrTyrArgProGlyAlaAsnLeuSer

850 870 890
 35 CTCTCCTGCTATGCAGCCTCTAACCCACCTGCACAGTACTCCTGGCTTATCAATGGACA
 LeuSerCysTyrAlaAlaSerAsnProProAlaGlnTyrSerTrpLeuIleAsnGlyThr

910 930 950
 40 TTCCAGCAAGGCACACAAGAGCTCTTTATCCCTAACATCACTGTGAATAATAGTGGATCC
 PheGlnGlnSerThrGlnGluLeuPheIleProAsnIleThrValAsnAsnSerGlySer

970 990 1010
 45 TATACCTGCCACGCCCAATAACTCAGTCACTGGCTGCCAACAGGACCACAGTCAAGACCATC
 TyrThrCysHisAlaAsnAsnSerValThrGlyCysAsnArgThrThrValLysThrIle

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1030 1050 1070
 5 ATAGTCACTGATAATGCTCTACACACAAGAAAAATGGCCTCTCACCTGGGGCCATTGCTGGC
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1090 1110 1130
 10 ATTGTGATTGGAGTAGTGGCCCTGGTTGCTCTGATAGCAGTAGCCCTGGCATGTTTTCTC
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1150 1170 1190
 15 CATTTCGGGAGACCGGCAGGGCAAGCGACCAGCGTGATCTCACAGAGCACAAACCCCTCA
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1210 1230 1250
 20 GTCTCCAAGCACACTCAGGACCACTCCAATGACCCACCTAACAAGATGAATGAAGTTACT
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1270 1290 1310
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1330 1350 1370
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1390 1410 1430
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 45 ATCTCCAGGCTGCCTGGTCACTGCCCCCTCTCTCAGTGTCAATAGATGAAAGTACATTGGG

1570 1590 1610
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 1690 1710 1730
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 GCCATAGCCCTTCAGGTTATGTCCTTTTCCATTAAGTACATGTGCCAGGAAACACCGAGAG
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 1810 1830 1850
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 1990 2010 2030
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 2050 2070 2090
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2230 2250 2270
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 2350 2370 2390
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 2590 2610 2630
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 35
 2650 2670 2690
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 2710 2730 2750
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 45
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 2830 2850 2870
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2890 2910 2930
TCTACACAGATGGAATCTCTTTGGGGCTAAGAGAAAGGTTTTATTCTATATTGCTTACCT
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2950 2970 2990
GATCTCATGTTAGGCCCTAAGAGGGCTTTCTCCAGGAGGATTAGCTTCGAGTTCTCTATACT
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3010 3030 3050
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ATGCTGTGCTGTGTTATTTAATTTTTCTGGCTAAGATCATGTCTGAATTATGTATGAAA
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3130 3150 3170
ATTATTCTATGTTTTTATAATAAAAATAATATATCAGACATCGAAAAAAAAAA,
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(2)

5 10 30 50
 CAGCCGTCCTCGAAGCGTTCTGGAGCCCAAGCTCTCCTCCACAGGTGAAGACAGGGCCA

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 15 130 150 170
 GGGCTTCTGCTCACAGCCTCACTTCTAACCCTTCTGGAACCCGCCCACCACTGCCCAGCTC
 GlyLeuLeuLeuThrAlaSerLeuLeuThrPheTrpAsnProProThrThrAlaGlnLeu

 20 190 210 230
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 25 250 270 290
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 30 310 330 350
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 35 370 390 410
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430 450 470
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5

490 510 530
CAGTTCCATGTATACCCGGAGCTGCCCAAGCCCTCCATCTCCAGCAACAACCTCCAACCCT
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550 570 590
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15 ValGluAspLysAspAlaValAlaPheThrCysGluProGluThrGlnAspThrThrTyr

610 630 650
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670 690 710
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730 750 770
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35 CCGGACACCCCCACCATTTCCCTTCAGACACCTATTACCGTCCAGGGGCAAACCTCAGC
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530 670 890
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910 930 950
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 40 GlyAsnThrThrLeuSerIleAsnProValLysArgGluAspAlaGlyThrTyrTrpCys

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 50
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1270 1290 1310
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1570 1590 1610
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1630
35 AAAAAAAAAA

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(3)

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110

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15

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130

150

170

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20

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190

210

230

ACTACTGAATCCATGCCATTCAATGTTGCAGAGGGGAAGGAGGTCTTCTCCTTGTCAC

25

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250

270

290

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30

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370

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410

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430

450

470

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55

490 510 530
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1150 1170 1190
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1270 1290 1310
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1330
 30 GAAAAAAAAAAAAAAAAA

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EP 0 346 710 B1

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	1981 gaataaacatgtaccacatttgcaaaaaa	2010

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(5)

1 ggggtggatccttaggctcatctccataggggagaaacacacatacagcagagaccatggga 59
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 1560 ttgcttttgcctctctatctgagtgccccccc 1591

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2. Véhicule de clonage recombinant apte à une réplication, comportant un produit d'insertion comprenant un acide nucléique selon la revendication 1.

25 3. Cellule qui a été transfectée, infectée par un véhicule de clonage recombinant selon la revendication 2, ou à laquelle on a injecté ce dernier.

4. Procédé pour préparer un polypeptide, ledit procédé comprenant les étapes consistant à :

(a) cultiver la cellule selon la revendication 3, et

30 (b) récupérer le polypeptide exprimé par ladite cellule.

5. Procédé pour préparer un anticorps dirigé contre un polypeptide, ledit procédé comprenant les étapes consistant à :

(a) préparer ledit polypeptide par le procédé selon la revendication 4,

35 (b) injecter ledit polypeptide dans un hôte capable de produire des anticorps, et

(c) récupérer lesdits anticorps.

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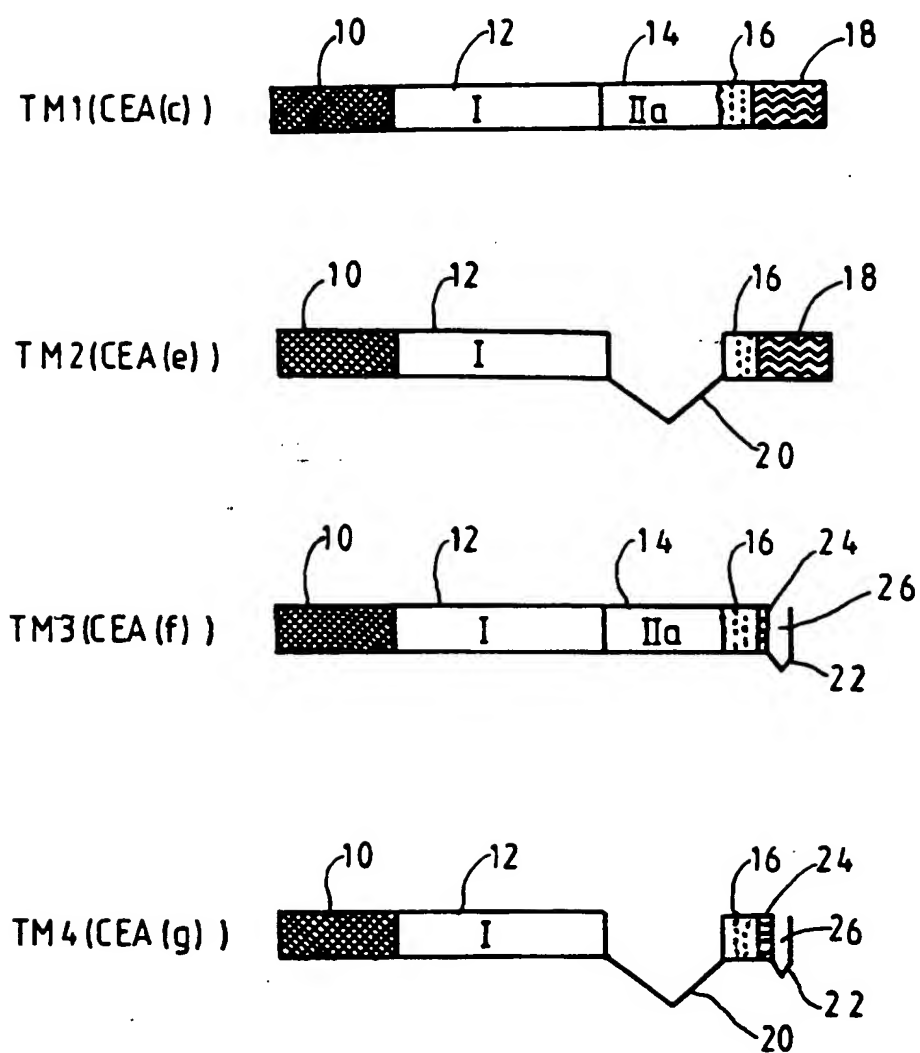


FIG.1